1	Use of rapid tests to predict quality traits of CIMMYT bread wheat genotypes grown
2	under different environments.
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14	For LWT - Food Science and Technology
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19 Abstract

20 At the International Maize and Wheat Improvement Center (CIMMYT), wheat quality improvement is an important goal of breeding. CIMMYT scientists develop germplasm, which is 21 diverse for quality traits intended for use in the preparation of different wheat-based products. 22 The integration of quality traits is complex due to the high cost of conducting traditional quality 23 24 tests. One option for tackling this problem is the use of such rapid-small-scale methods as Solvent Retention Capacity (SRC), SDS Sedimentation (SDSS) and Swelling Index of Glutenin 25 (SIG) to predict flour performance. The objectives of this study were to investigate the effect of 26 genotypes, contrasting environmental conditions and their interactions (GxE) on different rapid-27 28 small-scale tests, and to identify their suitability for use in prediction of quality traits. A significant GxE effect was observed for all three methodologies. Overall, SIG was found to be 29 the best predictor of gluten strength across different environments. It was also best at 30 31 determining bread-making quality in some environments, followed by SDSS for bread making. 32 SRC was found to be useful to select for gluten strength, but for extensibility and bread-making 33 more grain data is needed.

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Keywords: wheat quality; solvent retention capacity; SDS sedimentation; swelling index of
glutenin, alveograph; bread-making.

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41 **1. Introduction**

Successful adoption of new wheat varieties is largely dependent on the grain yield and 42 43 grain quality demands of average consumers and industrial food manufacturers (both semimechanized and mechanized industrial) in a given region. Due to these complex and multifaceted 44 needs, International Maize and Wheat Improvement Center (CIMMYT) scientists focus on the 45 46 core breeding challenges of simultaneous improvement of wheat production and quality for global distribution. With the estimated growth of the bakery industry at 6% globally, a need for 47 improved quality varieties has increased, but the integration of quality traits in a breeding 48 program remains a challenge. The focus is often on traits with more direct importance for 49 farmers such as grain yield or disease resistance. Additionally, high costs and time limitations 50 restrict the use of traditional quality tests conducted with the mixograph, farinograph, 51 52 alveograph, or end-use quality tests in large breeding programs where thousands of genotypes 53 are evaluated annually. Often there is not enough grain in early generations to conduct such 54 analyses. The absence of quality selection tests in the early or middle generations of a breeding 55 program could result in the development of advanced lines unsuitable for release due to related shortcomings of poor processing and end-use quality. 56

57 Small-scale, high-throughput methods for predicting flour performance, allow researchers 58 to make a broad selection, discard lines with insufficient quality, keeping those with improved 59 quality. The development of small scale dough testing equipment has been successful in several 60 cases (see Bekes, Lukow, Uthayakumaran, & Mann, 2003, for a good review). Several types of 61 equipment have been developed to work with small samples, including the two-gram mixograph 62 (standard mixographs use 35 g of flour) (Rath, Gras, Wrigley, & Walker, 1990), the micro Z-arm

mixer (4 g of flour), analogous to the farinograph (50-250 g of flour) (Haraszi, Gras, Tömösközi, 63 Salgó, & Bekes, 2004), which shows high correlations with standard equipment. Near infrared 64 65 (NIR) spectroscopy also has a great deal of potential to predict quality traits (Osborne, 2006), but is a costly, difficult for many breeding programs to afford. An economical and time saving 66 alternative is to use simple chemical tests, which result in correlated processing and end-use 67 68 quality traits. Sodium dodecyl sulfate sedimentation (SDSS), a commonly used traditional quality test, gives an overall idea of gluten quality and a fair prediction of bread-making 69 70 (Blackam & Gill, 1980; Peña, Amaya, Rajaram, & Mujeeb-Kazi, 1990). The Swelling Index of 71 Glutenin (SIG), developed by Wang and Kovacs (2002a), is a newer high throughput evaluation method, based on the same principle as SDSS (glutenin swelling capacity and insoluble glutenin 72 content) and has revealed the capacity to predict quality traits in bread wheat (Li, Wu, 73 Hernandez-Espinosa, & Peña, 2015; Wang and Kovacs 2002b). Finally, Solvent Retention 74 75 Capacity (SRC) is another significant means of measuring quality to determine which micro-76 methods have already been developed (Bettge, Morris, Demacon, & Kidwell, 2002; Guzman, Posadas-Romano, Hernandez-Espinosa, Morales-Dorantes, & Peña, 2015). SRC, originally 77 developed by Slade and Levine (1994), determines the capacity of flour to hold four solvents: 78 79 water, associated with the overall water holding capacity of all flours constituents; 50 g/L sodium carbonate, related to the damaged starch content of the flour; 500 g/L sucrose, 80 81 associated with the concentration of arabinoxylans; and 50 g/L lactic acid, associated with the 82 glutenin swelling capacity (Gaines, 2000). This method develops a flour-quality profile that 83 defines the contribution of individual grain components (Kweon, Slade, & Levine, 2011). This method has been widely used in soft wheats for cookie-making (Duyvejonck, Lagrain, Pareyt, 84 85 Courtin, & Delcour, 2011; Gaines (2004); Guttieri, Bowen, Gannon, Brien, & Souza, 2001;

86	Pasha, Anjum, & Butt, 2009; Zhang, Zhang, Zhang, He, & Peña, 2007) and in hard wheat
87	germplasm for other products (Colombo, Pérez, Ribotta, & León, 2008; Duyvejonck, Lagrain,
88	Dornez, Delcour, & Courtin, 2012; Li et al. 2015; Xiao, Park, Chung, Caley, & Seib, 2006).
89	However, most of the aforementioned studies, which used hard bread wheat were undertaken
90	with a limited number of genotypes and/or under a single set of environmental conditions. More
91	SRC data from diverse genetic backgrounds and environmental conditions are needed to validate
92	the value of this test in breeding programs and to understand its use relative to SDSS and SIG.
93	This study aimed mainly to investigate the effect of genotype (G), contrasting
94	environmental (E) conditions and their interactions (GxE) on SDSS, SIG and SRC. It also
95	aimed to identify the suitability of those methods for use in the prediction of quality traits in a set
96	of CIMMYT bread wheat lines grown worldwide.

98 2. Materials and methods

99 2.1 Plant Materials and Field Trials

A trial consisting of 54 CIMMYT bread wheat lines, including advanced lines, historical and 100 modern varieties (Electronic Supplementary Material 1), were sown in the 2012-2013 and 2013-101 2014 crop seasons in Ciudad Obregon (Mexico). The trial was set up in a lattice square design 102 with three replications and sown under six different environmental conditions: optimum 103 irrigation with drip (control environment); flat sown with basin irrigation; reduced irrigation or 104 moderate drought stress; severe drought stress; medium heat stress and severe heat stress. More 105 details of the trial are illustrated in Guzman, Autrique, Mondal, Singh, Govindan, Morales-106 107 Dorantes, et al. (2016)

109 *2.2 Grain and flour parameters*

110	Thousand kernel weight (g) and test weight (g/L) were evaluated with the digital image
111	system SeedCount SC5000 (Next Instruments, Condell Park, Australia). Grain protein (g/kg),
112	hardness (%) and moisture content were determined by near-infrared spectroscopy (NIR Systems
113	6500, Foss, Hillerød, Denmark) calibrated based on official American Association of Cereal
114	Chemists (AACC) methods 39-10 and 46-11A (AACC, 2010). Grain samples previously
115	conditioned at 140-160 (g/kg) of moisture were milled into flour using Brabender Quadrumat Jr
116	(C. W. Brabender OHG, Duisburg, Germany).
117	Measurement of SDSS volume was carried out according to Peña et al. (1990). SIG was
118	determined with lactic acid according to the second variant of the method used by Wang and
119	Kovacs (2002a). SRC was carried out according to Guzman at al. (2015) with four solvents:
120	water, sodium carbonate, sucrose and lactic acid. All data from these tests are available in
121	Electronic Supplementary Material 1.
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123	2.3 Rheological and baking tests
124	Dough development properties were determined by Mixograph of Swanson (National

125 Mfg., Lincoln, U.S.A.) using 35 g of flour (AACC method 54-40A), obtaining dough

development time and %Torque*min. The Chopin Alveograph (Trippette & Renaud, Villeneuve-

127 la-Garenne, France) was used to determine dough tenacity, extensibility, strength (ALVW) and

tenacity/extensibility ratio (ALVP/L) (AACC 54-30A) using 60 g of flour. The bread-making

129 process was conducted using the direct dough method (AACC method 10-09) and bread loaf

130 volume was determined by rapeseed displacement using a volumeter.

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132 2.4 Statistical Analysis

Pearson correlation coefficients (r) and the significance of each comparison in the study wereobtained using SAS.

Combined analyses of variance (ANOVA) across environments for grain and other quality
traits were performed using procedure Proc Anova of the SAS statistical software (SAS, 2014).
The means of genotypes in each environment throughout the two-year period during which
the trials were undertaken were used in the variable selection stepwise procedure using an alpha
level of 0.0001 (Proc Stepwise, SAS version 9.4, 2014). All multiple regression equations are
detailed in Electronic Supplementary Material 2.

141 **3. Results and discussion**

142 *3.1 Grain and flour characteristics*

The data of both cropping cycles were quite similar (data not shown), explained by the high 143 heritability revealed in all traits (Table 1). A wide range in grain characteristics was observed in 144 145 genotypes across different environments. Test weight and particularly thousand kernel weight grain morphology parameters showed great variability, between and within each environment. 146 The range of values for grain hardness was somewhat smaller (32-55%), without any samples 147 showing real soft texture (>55%). For grain protein content, the variation was also important 148 149 (107-175 g/kg) and larger in such highly stressed environments as severe drought stress or severe 150 heat stress. Compared to the optimum environment (110-141 g/kg) protein content was high in severe drought (12-17.1%) and heat stress environments (121-175 g/kg). Across environments, 151 152 test weight and thousand kernel weight showed a negative association with grain protein content, r = -0.48 and -0.52, respectively, (p<0.0001), due to a dilution or concentration effect depending 153 on grain size. In SRC tests, lactic acid SRC showed the highest variation and the lowest was 154

shown in sodium carbonate SRC, with water SRC and sucrose SRC (also showing smaller ranges 155 than lactic acid SRC. The range of lactic acid SRC in control environment (105-162.3%) was 156 157 similar to that found by Duyjevonck et al. (2012), Li et al. (2015) and Xiao et al. (2006) (studies conducted using hard wheat). Lactic acid SRC is related to gluten strength (Gaines 2000) and 158 thus higher values are expected as this study included hard or semi-hard bread wheat lines often 159 160 used for products that require medium-strong gluten compared to soft wheat or cookie-making that requires weak gluten content (Guttieri et al. 2001; Zhang et al. 2007). Lactic acid SRC 161 162 significantly increased in drought-stressed environments, most likely due to the increase in grain 163 protein content (r value between lactic acid SRC and grain protein content in the whole trial was 0.33, p<0.0001), although no such effect was seen in medium and severe heat-stress 164 environments. The increase in grain protein content in heat-stressed environments is done 165 together with qualitative change in protein composition (a decrease in glutenin-to-gliadin ratio) 166 that influenced the lactic acid SRC values (and weaker gluten). Previous studies have reported 167 168 such observations (Blumenthal, Bekes, Gras, Barlow, & Wrigley, 1995). A similar trend was found also in SDSS and SIG. A significant correlation was observed between lactic acid SRC 169 and SDSS and SIG (r = 0.32 and 0.48, respectively, p<0.0001), and SDSS with SIG (r = 0.76, 170 171 p<0.0001). The same fact was observed by Duyvejonck et al. (2012) with Zeleny sedimentation and lactic acid SRC. 172 173 Sodium carbonate SRC is related with grain hardness (r = -0.55, p<0.0001) because sodium

carbonate SRC is related to flour starch damage. Both high and low sodium carbonate SRC
values were observed, higher values indicating hard texture (Duyvejonck et al. 2012; Xiao et al.
2006) and lower values soft texture (Bettge et al. 2002; Guttieri, Souza, & Sneller, 2008). Higher
sodium carbonate SRC values were found in drought environments, associated to lower grain

178	hardness values (hard grain), and lower in severe heat-stress environment, associated to softer
179	texture. The same trend was found in sucrose SRC, which is related to the pentosans and
180	somewhat to gliadin content, and water SRC (Gaines 2000). SRC, SDSS and SIG values were
181	affected by specific irrigation treatments and temperature regimes similar to those reported by
182	Walker, Campbell, Carter, & Kidwell (2008), Zhang et al. (2007) and in contrast to results
183	reported by Guttieri, McLean, Lanning, Talbert, & Souza (2002). As grain protein content,
184	sucrose SRC showed an inverse correlation with test weight and thousand kernel weight ($r = -$
185	0.22 and -0.31, respectively, p<0.0001), probably because pentosans are located in the cell wall,
186	which is more concentrated in the grain.
187	Therefore, the samples appear to represent a wide spectrum of grain and quality traits. This
188	was confirmed with further rheological analysis (mixograph, alveograph and bread-making)

(data not showed). 189

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191 3.2 Analysis of variance

For any methodology to be suitable for use as a wheat-quality improvement selection tool, it 192 must detect significant differences among genotypes and have low GxE effects for consistent 193 194 selection. This has been already studied for SRC in soft wheat, with several studies reporting 195 high genotypic and limited or no significant GxE effect (Guttieri and Souza 2003; Guttieri et al. 196 2001, 2002; Pasha et al. 2009). In contrast, significant GxE effects were found in a study by 197 Walker et al. (2008) and by Zhang, Zhang, & He, (2008) in a study involving soft spring and winter wheat lines grown in different locations of Washington state in the United States. 198 199 In the current study, genotype was the most important source of variation (Table 2) for water 200 SRC, sodium carbonate SRC, sucrose SRC and SDSS, and the second one, lactic acid SRC and

SIG. This is most likely due to the large diversity for quality traits that the set of lines used had 201 and for the high heritability of those traits. Environmental effect was significant, particularly for 202 203 lactic acid SRC and SIG, explaining more than 40% of the variation found in those traits. This strong environmental effect was due to highly contrasting field management conditions used in 204 205 each trial, particularly in severe drought and heat-stress environments. The result for lactic acid 206 SRC is in contrast to that observed by Guttieri et al. (2001, 2002) where no significant environmental effect was observed. On the contrary, Walker et al. (2008) found a significant 207 208 environmental effect on spring wheat lactic acid SRC values. A significant environmental effect 209 implies that for accurate measurements and efficient selection of genotypes with the rapid tests, control genotypes must be evaluated in diverse environments. This applies to SIG and SDSS 210 tests as well, based on the results in this study. The strong environmental effect on the three traits 211 was somewhat expected as all of them depend on some way in protein content, which is well 212 known to be highly influenced by different environmental conditions. 213 214 The year effect was minor. All the interactions related with the genotype (GxE, GxY and GxExY) were highly significant and together explained around 12-19% of the variation, except 215 for lactic acid SRC (9%). This result agrees with Walker et al. (2008), who also found significant 216 217 interactions for SRC tests in samples produced in a wide range of environments. Previously, Guttieri et al. (2001, 2002) did not find significant GxE for SRC tests. In our study, although the 218 219 variation explained by interaction effects is low, their significance serves as a recommendation 220 that multiple environments should be used for selection. This approach should be used in 221 breeding programs with a multiregional or global focus similar to CIMMYT's, which involves 222 the use of contrasting growing conditions in different environments and the use of diverse

223 germplasm that could potentially adapt to each of them.

Among lactic acid SRC, SDSS and SIG, SDSS showed stronger genetic control, thereby suggesting its efficiency in selection for better quality genotypes in breeding programs. Other SRC solvents (water, sodium carbonate, sucrose) also had high genotype effect, making them suitable for selection, although high GxE interactions reveal the need for selection in multiple environments.

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0 *3.3 Prediction models for processing and end-use quality traits*

231 In breeding for wheat quality, there are other traits that are taken into consideration in 232 addition to bread-making while evaluating lines, which means that flour quality cannot be expressed by a single trait, but depends on the interaction of several factors. Among them, the 233 most important are dough strength and extensibility, which can be measured by the mixograph 234 (dough development time and Torque) and by the alveograph (balance between tenacity and 235 extensibility, ALVP/L, gluten strength, ALVW). The above mentioned traits together with grain 236 237 hardness are used in the CIMMYT breeding program to classify the lines evaluated in five different end-use types (Guzman, Medina-Larqué, Velu, González-Santoyo, Singh, Huerta-238 Espino, & Peña, 2014). Well-trained operators and a large quantity of flour are required for the 239 240 aforementioned tests. Therefore, the development of simple tests that use just a few grams of flour is essential for early generation testing in breeding programs. Stepwise multiple regression 241 242 models were deployed using six different sets of wheat grain and flour quality parameters. The 243 first set included data from SRC tests carried out with the four different solvents; second and 244 third sets only included the data from SDSS and SIG, respectively. The next sets (fourth, fifth 245 and sixth) were equivalent to the first three tests, but in this case data of test weight, thousand 246 kernel weight, grain hardness and grain protein content were added to all of them. Those

parameters are routinely and easily obtained with such high-throughput methodologies as image 247 analysis and NIR (Osborne, 2006), which require a small amount of grain. M aking different sets 248 249 to build prediction models was intended to use as few parameters as possible to really save resources and time, so that only highly significant traits (p<0.0001) to each model were included. 250 SRC, SDSS and SIG were not combined in the same set to make clear which of the three popular 251 252 tests is the best option for analysis of lines from the breeding program. Regression models were 253 first built based on all environmental data of the trial and subsequently for each specific 254 environment (data from both years). The r values of each model to predict different quality traits 255 are shown in Table 3. In Electronic Supplementary Material 2 all multiple regression equations are included. 256

From the first three sets of parameters (SRC, SDSS or SIG) we obtained models that 257 explained much of the variation in target traits. In models developed with all trial data, SIG was 258 259 found to be the best predictor of gluten strength, with r values of 0.81 and 0.90 for torque and 260 alveograph gluten strength (ALVW), respectively. SRC (in this case only lactic acid SRC as the other solvents were not found significant for prediction; see Electronic Supplementary Material 261 2) was also a very good predictor for those traits (r = 0.85 for ALVW) followed by SDSS (r =262 263 0.77 for ALVW). To predict alveograph gluten tenacity and extensibility, the models built with both SDSS and SIG were unable (not significant) to predict the important alveograph 264 265 tenacity/extensibility ratio (ALVP/L) value. Predictions with lactic acid SRC for ALVP/L 266 improved slightly ALVP/L (r = 0.58) although they were far from accurate. For alveograph 267 tenacity and alveograph extensibility, the combination of sodium carbonate SRC and water SRC, 268 respectively, with lactic acid SRC, gave acceptable values of prediction. When predicted values 269 of alveograph tenacity and alveograph extensibility were obtained and used to calculate

ALVP/L, the prediction of this trait is improved (r = 0.64). For bread-making, SIG and SDSS (r0.67 and 0.66) were remarkably better than lactic acid SRC (0.51). Overall, trial data revealed that SIG was the most useful test to predict end-use quality and gluten strength, and for selection of gluten extensibility lactic acid SRC, sodium carbonate SRC and water SRC should be carried out.

275 The next research priority was to determine if models built with data from a specific 276 environment would have the same potential to predict quality traits in that environment. In 277 environments with better field conditions lactic acid SRC and SIG were both excellent predictors 278 of gluten strength, being lactic acid SRC slightly better. SDSS was also acceptable but did not reach higher r than 0.67 for ALVW. In drought-stressed environments results were the same, but 279 SDSS showed better prediction than previously in medium drought stress environments (r =280 (0.78) and something less in severe drought stress environments (r = 0.73). In heat-stressed 281 environments, lactic acid SRC lost some prediction power but water SRC gained significantly in 282 283 medium heat-stress environments to reach SRC r value of 0.74 and the same in severe heat-stress environments, where of all SRC solvents only lactic acid SRC again played a significant role. 284 SDSS predictions for gluten strength were improved in medium heat-stress environments and 285 286 even more in severe heat stress environment, but SIG fared better (r = 0.84). Again, results indicated that SIG is the best rapid test to predict gluten strength consistently across different 287 288 environments. Wang and Kovacs (2002b), and Li et al. (2015) reached the same conclusions 289 after conducting similar experiments. The ability of SIG to predict gluten strength compared to 290 other tests is due to its higher association with the insoluble glutenin fraction in flour (Wang and 291 Kovacs 2002a).

For alveograph tenacity/extensibility ratio (ALVP/L) prediction SIG and SDS models 292 results were completely insignificant. Lactic acid SRC data was also insignificant. This could be 293 294 due to the interaction of other factors, apart from the insoluble glutenin fraction, which affect ALVP/L, reflecting the balance between dough tenacity and extensibility. However, other SRC 295 solvents had a certain amount of prediction power. For example, sodium carbonate SRC (r =296 297 0.61 and 0.68) revealed this characteristic when sown flat with basin irrigation and medium heat-298 stress environments, respectively. Duyvejonck et al. (2012) also observed the association of 299 sodium carbonate SRC with alveograph tenacity. Dexter et al. (1994) observed that starch with 300 higher damage resulted in more water retention by flour, leading to stiffer dough and related increased dough resistance. Although the r values were moderate, experiments showed they may 301 not lead to accurate prediction of ALVP/L. 302

For bread-making predictions, SIG was found to be the best in most of the environments 303 (r not higher than 0.64), excluding flat sown with basin irrigation and severe heat-stress 304 305 environments were SDSS was the best one (r = 0.64 and 0.72, respectively). Lactic acid SRC was only near SDSS and SIG in predicting bread loaf volume in the control environment, so that 306 test is not highly recommended to predict loaf volume if it is not complemented with other data. 307 308 This finding is in disagreement with Xiao et al. (2006), who found lactic acid SRC better than SDSS when predicting loaf volume (r of 0.83 vs. 0.76) and with Colombo et al. (2008) (r of 0.72 309 310 vs. 0.51). Study results are in partial agreement with Li et al. (2015), who showed SDSS to be 311 the best predictor for bread loaf volume sown flat with basin irrigation, severe drought and heat-312 stress enviroments, in comparison to lactic acid SRC or SIG. Wang and Kovacs (2002b) found 313 SIG and SDSS of equal benefit to predicting bread loaf volume (r = 0.54).

When additional test weight, thousand kernel weight, grain hardness and grain protein 314 content data was added to the original sets of SRC, SDSS and SIG data, the predictions were 315 significantly increased for some traits and in some specific environments. For gluten strength 316 traits, overall predictions were not increased to a great degree, although in some cases progress 317 was evident. For example, SDS + grain hardness in full irrigation environments or lactic acid 318 319 SRC + thousand kernel weight in severe heat stress environment. SIG alone or with such other parameters as grain protein content in full irrigation environments, continued being the most 320 321 useful trait to predict gluten strength. With regard to ALVP/L, in most circumstances new data 322 added to the models did not involve an increase in the prediction with SDSS or SIG. Therefore, both rapid tests should be discarded if the objective is to make selection for gluten extensibility. 323 In the case of SRC, a significant increase in prediction for ALVP/L was achieved when other 324 traits were added to the model. In the model showing all data, lactic acid SRC + sodium 325 326 carbonate SRC+ test weight explained 65% of the ALVP/L variation. In the control environment, 327 the use of lactic acid RC + sodium carbonate SRC + grain protein content resulted in r of 0.61, in areas flat sown with basin irrigation environment lactic acid RC + grain protein content + test 328 weight resulted in r of 0.7, while in severe heat stress environment r of 0.6 (and of 0.69 if 329 330 ALVP/L is manually calculated from predicted alveograph tenacity and extensibility) was obtained with water SRC + thousand kernel weight. Those values, although not very high, could 331 332 be sufficient to undertake a broad selection for the purpose of discarding tenacious breeding 333 lines. Additionally, in medium heat stress environment the prediction for ALVP/L reached r of 0.78 (sodium carbonate SRC + test weight + grain protein content), which was sufficient to make 334 a more accurate selection. 335

Finally, for bread-loaf volume prediction, the addition in the model of more grain traits 336 data resulted in very important increases, particularly for SRC. Different combinations of two 337 338 solvents (lactic acid SRC + sodium carbonate SRC or sucrose SRC) + grain protein content + test weight or thousand kernel weight led to predictions with r of 0.78 in full irrigation 339 environments, higher than those of SDSS or SIG in combination with other grain traits. For 340 341 drought-stressed environments, better predictions are given by SIG + thousand kernel weight, but similar results are obtained with SRC or SDSS. For heat-stressed environments, SDS in 342 343 combination with grain protein content + thousand kernel weight or with test weight had the 344 higher values. Therefore, for bread making there are various choices of rapid tests - some better than others – depending on the environment and the availability of other grain traits. It seems 345 unsuitable to select only one rapid test for all the environments, but if the priority of the breeding 346 program is to improve gluten strength and bread-making, research shows that SIG is overall 347 probably the best test. The results from our models for predicting loaf volume are slightly 348 349 inferior to those from Xiao et al. (2006) using similar sets of data (r of 0.83 or 0.87), probably because they also include milling and mixograph parameters in the model. Using those 350 parameters, lactic acid SRC, combined with other traits, produced better result than SDSS 351 352 combined with other traits, which for this study occurred in full irrigation environments. As expected, grain protein content data was incorporated as a significant factor into 353 354 several of the predictions models, as it is well known that processing and end-use quality traits 355 are highly dependent on protein quality and quantity. Test weight and thousand kernel weight 356 were significant in some models because they explained the concentration or dilution of other 357 grain components that affect quality. Grain hardness was also significant in some models due to

its profound effect on dough water absorption, which can influence several rheological and end-use quality tests.

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361 **4.** Conclusions

In the current study, a large and diverse set of hard bread wheat germplasm was analyzed 362 363 with different rapid tests. Research confirmed that the three methodologies are under strong genetic control, although environmental effects and GxE interactions were significant in all cases 364 with an impact significant enough to consider the necessity of multi-location evaluation. The 365 366 prediction efficiency for several quality traits of each rapid test was evaluated in different environments and in combination with other grain traits, an important component to help 367 breeding programs select the best methodology depending on field conditions and traits of 368 interest. In addition to these discoveries, when choosing a rapid-small-scale test to screen for 369 quality attributes in a breeding program, research revealed the importance of considering the 370 371 number of samples that can be analyzed per unit time, the cost of that activity and the equipment required to do it. In this regard, SRC tests and SIG, performed as described by Guzman et al. 372 (2015) and Wang and Kovacs (2002a) require more expensive equipment (thermomixer and 373 374 centrifuge for small test tubes), which also ensure high repetitiveness of the analysis done. Besides, both methodologies imply the use of Eppendorf test tubes, which could be discarded 375 376 after running the test or be cleaned, although in the case of SRC this is extremely time-377 consuming because the pellets remain strongly adhered to the bottom of the tube. The SDSS 378 performed as described by Peña et al. (1990) requires 25 ml test tubes that need to be cleaned 379 after the test. The three methodologies are high-throughput and hundreds of samples can be 380 analyzed in one day. SDSS is faster because it requires a higher flour amount (1g), making the

381	sample weighting process much faster than for LARC or SIG, for which 0.3g and 0.02g must be
382	accurately weighted in a precision balance. Evaluating these methodological characteristics is
383	also important when selecting a methodology to be implemented in the breeding program.
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390	
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Table 1. Mean values, value ranges and heritability for grain traits and rapid tests found for 54 bread wheat genotypes in the whole and in each environment of the trial.

	Total			Full drip irrigation Full basin irrigation			Mild drought stress Severe drought stress			Mild heat stress		Severe heat stress			
	Mean	Range	Heritability	Mean	Range	Mean	Range	Mean	Range	Mean	Range	Mean	Range	Mean	Range
Grain traits Test weight (g/L) Thousand	802	679-846	0.88	812	767-832	811	756-841	815	751-842	800	742-828	802	744-830	773	679-807
kernel weight (g) Grain	41.6	23.5-59.3	0.97	46.5	29.4-56.7	43.4	28.1-55.1	45.8	30.6-57.6	37.6	26.7-52.5	42.6	30.8-59.3	33.3	23.5-45.9
hardness (%)	42.6	32-55	0.9	42.2	36-50.8	43.2	36-52.4	40.0	35.0-48.8	41.5	32-51	43.3	36-53.4	45.5	38-55
Grain protein content (g/kg)	135	107-175	0.83	124	110-141	120	107-138	137	117-158	150	120-171	129	114-152	150	121-175
Rapid tests Water SRC (%)	70.2	56.4-83	0.92	69.8	58.2-78	68.8	56.4-77.3	72.1	61.5-83.1	72.8	63.3-81.4	69.8	60.5-77.3	68.0	57.3-77
Sodium carbonate SRC (%)	79.1	62.4-94.3	0.94	79.5	67.1-90	78.2	65.5-88.6	81.4	69.2-94.3	81.1	69.1-89.4	78.4	67-89.7	75.9	62.4-86.5
Sucrose SRC (%)	94.1	80.3-110.6	0.92	91.4	81.2-101.7	89.9	80.3-98.5	96.4	82.2-109.8	98.9	88.7-110.7	92.9	83.9-105.5	95.3	82.4-108-8
Lactic acid SRC (%)	128.4	87.4-196.2	0.95	125.6	105-162.3	122.2	92.7-163.1	139.1	108.4-177.5	146.9	119.6-196.2	120.5	91.8-160.9	116.3	87.4-152.2
SDSS (ml)	15.7	8-23.0	0.96	13.9	8-21.5	13.8	8-20.5	15.7	10-21.5	18.7	11.5-23	15.0	9-22.5	17.1	9-22.0
SIG	5.9	4.2-7.9	0.92	5.6	4.6-6.6	5.5	4.2-6.5	6.0	5-7.4	6.6	5.1-7.9	5.7	4.7-7.3	6.1	4.9-7.4

SRC: solvent retention capacity; SDSS: SDS sedimentation; SIG: swelling index of glutenin.

•	Sodium								Lactic acid						
		SDSS		Water SRC		carbonate SRC		Sucrose SRC		SRC		SIG			
	DF	SS	% SS	SS	% SS	SS	% SS	SS	% SS	SS	% SS	SS	% SS		
Genotype	53	7861	54	8411	48	13934	54	15295	41	125491	40	145	34		
Environment	5	4024	28	3711	21	4570	18	12075	33	152620	49	178	41		
Year	1	15	0.1	177	1	62	0.2	74	0.2	7	0**	23	5		
GxE	265	948	6	1961	11	2587	10	4257	12	15018	5	30	7		
GxY	53	211	1	265	1	504	2	433	1	2843	1	6	1		
YxE	5	607	4	1152	6	1545	6	1355	4	4026	1	10	2		
ExGxY	265	669	5	1057	6	1592	6	1664	4	9434	3	25	6		

Table 2. Effects of genotype, environment and year and their interactions on the rapid tests. Sum of squares and percentages of sum of squares respect to the total sum of squares obtained from ANOVA analysis are showed.

*All the effects were highly significant (p<0.0001), except **.

SDSS: SDS sedimentation; SRC: solvent retention capacity; SIG: swelling index of glutenin.

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Table 3. r correlation values obtained with different multiple regression equations to predict processing and end-us
quality traits using rapid tests and grain traits in the whole trial and in each environment.

Total (six environments)	Dough development time	Torque	Alveograph W	Alveograph P/L	Alveograph tenacity	Alveograph extensibility	Alveograph P/L*	Loaf volume
Set 1: SRC	0.63	0.74	0.85	0.58	0.78	0.70	0.64	0.51
Set 2: SDSS	0.66	0.73	0.77	NS	NS	0.62	NS	0.66
Set 3: SIG	0.73	0.82	0.91	NS	0.52	0.52	NS	0.68
Set 4: SRC + GT	0.63	0.74	0.85	0.64	0.81	0.77	0.69	0.78
Set 5: SDS + GT	0.66	0.73	0.79	NS	NS	0.71	0.51	0.74
Set 6: SIG + GT	0.73	0.82	0.91	0.48	0.65	0.69	0.51	0.80
Full drip irrigation	Dough development	Torque	Alveograph W	Alveograph P/L	Alveograph tenacity	Alveograph extensibility	Alveograph P/L*	Loaf volume

	time							
Set 1: SRC	0.60	0.72	0.83	0.50	0.71	0.57	0.52	0.58
Set 2: SDS	0.60	0.65	0.64	NS	NS	0.57	NS	0.57
Set 3: SIG	0.62	0.71	0.79	NS	0.48	0.52	NS	0.59
Set 4: SRC + GT	0.60	0.72	0.83	0.62	0.77	0.62	0.58	0.78
Set 5: SDS + GT	0.64	0.68	0.73	NS	NS	0.64	0.48	0.73
Set 6: SIG + GT	0.68	0.75	0.81	0.47	0.65	0.65	0.50	0.74
Full basin irrigation	Dough development time	Torque	Alveograph W	Alveograph P/L	Alveograph tenacity	Alveograph extensibility	Alveograph P/L*	Loaf volume
Set 1: SRC	0.67	0.76	0.84	0.62	0.77	0.67	0.68	0.57
Set 2: SDS	0.61	0.64	0.68	NS	NS	0.64	NS	0.65
Set 3: SIG	0.67	0.73	0.82	NS	0.52	NS	NS	0.58
Set 4: SRC + GT	0.76	0.82	0.85	0.70	0.77	0.84	0.71	0.81
Set 5: SDS + GT	0.71	0.75	0.73	0.57	0.57	0.77	0.55	0.77
Set 6: SIG + GT	0.76	0.73	0.84	0.57	0.71	0.73	0.59	0.75
Mild drought stross	Dough development	Torque	Alveograph W	Alveograph P/I	Alveograph	Alveograph	Alveograph P/I *	Loaf
Mild drought stress	Dough development time	Torque	Alveograph W	Alveograph P/L	Alveograph tenacity	Alveograph extensibility	Alveograph P/L*	Loaf volume
Mild drought stress Set 1: SRC Set 2: SDS	Dough development time 0.66 0.66	Torque 0.77 0.73	Alveograph W 0.85 0.78	Alveograph P/L 0.50 NS	Alveograph tenacity 0.67 NS	Alveograph extensibility 0.66 0.62	Alveograph P/L* 0.57 NS	Loaf volume 0.61
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG	Dough development time 0.66 0.66 0.66	Torque 0.77 0.73 0.77	Alveograph W 0.85 0.78 0.85	Alveograph P/L 0.50 NS NS	Alveograph tenacity 0.67 NS NS	Alveograph extensibility 0.66 0.62 0.52	Alveograph P/L* 0.57 NS NS	Loaf volume 0.61 0.64 0.65
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT	Dough development time 0.66 0.66 0.66 0.66	Torque 0.77 0.73 0.77 0.77	Alveograph W 0.85 0.78 0.85 0.85	Alveograph P/L 0.50 NS NS 0.50	Alveograph tenacity 0.67 NS NS 0.67	Alveograph extensibility 0.66 0.62 0.52 0.66	Alveograph P/L* 0.57 NS NS 0.57	Loaf volume 0.61 0.64 0.65 0.71
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT Set 5: SDS + GT	Dough development time 0.66 0.66 0.66 0.66 0.66	Torque 0.77 0.73 0.77 0.77 0.73	Alveograph W 0.85 0.78 0.85 0.85 0.85 0.78	Alveograph P/L 0.50 NS NS 0.50 NS	Alveograph tenacity 0.67 NS NS 0.67 NS	Alveograph extensibility 0.66 0.62 0.52 0.66 0.66	Alveograph P/L* 0.57 NS NS 0.57 NS	Loaf volume 0.61 0.64 0.65 0.71 0.71
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT Set 5: SDS + GT Set 6: SIG + GT	Dough development time 0.66 0.66 0.66 0.66 0.66 0.66	Torque 0.77 0.73 0.77 0.77 0.73 0.73 0.77	Alveograph W 0.85 0.78 0.85 0.85 0.78 0.78 0.85	Alveograph P/L 0.50 NS NS 0.50 NS NS	Alveograph tenacity 0.67 NS NS 0.67 NS NS	Alveograph extensibility 0.66 0.62 0.52 0.66 0.66 0.66	Alveograph P/L* 0.57 NS NS 0.57 NS NS	Loaf volume 0.61 0.64 0.65 0.71 0.71 0.71
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT Set 5: SDS + GT Set 6: SIG + GT	Dough development time 0.66 0.66 0.66 0.66 0.66 0.66 0.66 Dough development	Torque 0.77 0.73 0.77 0.77 0.77 0.73 0.77	Alveograph W 0.85 0.78 0.85 0.85 0.78 0.85 0.78 0.85 Alveograph	Alveograph P/L 0.50 NS 0.50 NS 0.50 NS NS Alveograph	Alveograph tenacity 0.67 NS 0.67 NS 0.67 NS NS Alveograph	Alveograph extensibility 0.66 0.62 0.52 0.66 0.66 0.66 0.62 Alveograph	Alveograph P/L* 0.57 NS 0.57 NS 0.57 NS NS NS	Loaf volume 0.61 0.64 0.65 0.71 0.71 0.71 0.71 Loaf
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT Set 5: SDS + GT Set 6: SIG + GT Severe drought stress	Dough development time 0.66 0.66 0.66 0.66 0.66 0.66 0.66 0.6	Torque 0.77 0.73 0.77 0.77 0.73 0.77 D.77	Alveograph W 0.85 0.78 0.85 0.85 0.78 0.85 0.78 0.85 Alveograph W	Alveograph P/L 0.50 NS 0.50 NS 0.50 NS NS Alveograph P/L	Alveograph tenacity 0.67 NS 0.67 NS 0.67 NS NS Alveograph tenacity	Alveograph extensibility 0.66 0.62 0.52 0.66 0.66 0.66 0.62 Alveograph extensibility	Alveograph P/L* 0.57 NS NS 0.57 NS NS NS Alveograph P/L*	Loaf volume 0.61 0.64 0.65 0.71 0.71 0.71 0.71 Loaf volume
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT Set 5: SDS + GT Set 6: SIG + GT Severe drought stress Set 1: SRC	Dough development time 0.66 0.66 0.66 0.66 0.66 0.66 Dough development time 0.68	Torque 0.77 0.73 0.77 0.77 0.73 0.77 D.73 0.77 D.73	Alveograph W 0.85 0.78 0.85 0.85 0.78 0.85 0.85 Alveograph W 0.86	Alveograph P/L 0.50 NS 0.50 NS 0.50 NS NS Alveograph P/L NS	Alveograph tenacity 0.67 NS 0.67 NS 0.67 NS NS Alveograph tenacity 0.62	Alveograph extensibility 0.66 0.62 0.52 0.66 0.66 0.62 Alveograph extensibility 0.56	Alveograph P/L* 0.57 NS 0.57 NS 0.57 NS NS Alveograph P/L* 0.45	Loaf volume 0.61 0.64 0.65 0.71 0.71 0.71 0.71 Loaf volume 0.49
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT Set 5: SDS + GT Set 6: SIG + GT Severe drought stress Set 1: SRC Set 2: SDS	Dough development time 0.66 0.66 0.66 0.66 0.66 0.66 0.66 Dough development time 0.68 0.63	Torque 0.77 0.73 0.77 0.77 0.73 0.77 D.77 0.73 0.77 Torque 0.78 0.69	Alveograph W 0.85 0.78 0.85 0.85 0.78 0.85 0.78 0.85 Alveograph W 0.86 0.73	Alveograph P/L 0.50 NS 0.50 NS 0.50 NS NS Alveograph P/L NS NS	Alveograph tenacity 0.67 NS 0.67 NS NS NS Alveograph tenacity 0.62 NS	Alveograph extensibility 0.66 0.62 0.52 0.66 0.66 0.62 Alveograph extensibility 0.56 NS	Alveograph P/L* 0.57 NS 0.57 NS 0.57 NS NS Alveograph P/L* 0.45 NS	Loaf volume 0.61 0.64 0.65 0.71 0.71 0.71 0.71 Loaf volume 0.49 0.58
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT Set 5: SDS + GT Set 6: SIG + GT Set 1: SRC Set 1: SRC Set 2: SDS Set 3: SIG	Dough development time 0.66 0.66 0.66 0.66 0.66 0.66 0.66 Dough development time 0.68 0.63 0.75	Torque 0.77 0.73 0.77 0.77 0.73 0.77 D.73 0.77 D.78 0.69 0.82	Alveograph W 0.85 0.78 0.85 0.85 0.78 0.85 Alveograph W 0.86 0.73 0.89	Alveograph P/L 0.50 NS 0.50 NS 0.50 NS NS Alveograph P/L NS NS NS	Alveograph tenacity 0.67 NS 0.67 NS NS NS Alveograph tenacity 0.62 NS NS	Alveograph extensibility 0.66 0.62 0.52 0.66 0.66 0.62 Alveograph extensibility 0.56 NS NS	Alveograph P/L* 0.57 NS 0.57 NS 0.57 NS NS Alveograph P/L* 0.45 NS NS	Loaf volume 0.61 0.64 0.65 0.71 0.71 0.71 0.71 Uoaf volume 0.49 0.58 0.65
Mild drought stress Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT Set 5: SDS + GT Set 6: SIG + GT Set 1: SRC Set 2: SDS Set 3: SIG Set 4: SRC + GT	Dough development time 0.66 0.66 0.66 0.66 0.66 0.66 0.66 Dough development time 0.68 0.63 0.75 0.68	Torque 0.77 0.73 0.77 0.77 0.73 0.77 0.73 0.77 Torque 0.78 0.69 0.82 0.78	Alveograph W 0.85 0.78 0.85 0.85 0.78 0.85 Alveograph W 0.86 0.73 0.89 0.86	Alveograph P/L 0.50 NS 0.50 NS 0.50 NS NS Alveograph P/L NS NS NS NS	Alveograph tenacity 0.67 NS 0.67 NS 0.67 NS NS Alveograph tenacity 0.62 NS NS 0.71	Alveograph extensibility 0.66 0.62 0.52 0.66 0.66 0.62 Alveograph extensibility 0.56 NS NS NS	Alveograph P/L* 0.57 NS NS 0.57 NS NS Alveograph P/L* 0.45 NS NS NS	Loaf volume 0.61 0.64 0.65 0.71 0.71 0.71 0.71 Loaf volume 0.49 0.58 0.65 0.71

Set 6: SIG + GT	0.59	0.69	0.75	NS	NS	NS	NS	0.75
Mild heat stress	Dough development time	Torque	Alveograph W	Alveograph P/L	Alveograph tenacity	Alveograph extensibility	Alveograph P/L**	Loaf volume
Set 1: SRC	0.63	0.69	0.75	0.69	0.76	0.66	0.71	0.45
Set 2: SDS	0.62	0.62	0.75	NS	NS	0.58	NS	0.59
Set 3: SIG	0.59	0.69	0.75	NS*	NS	NS	NS	0.65
Set 4: SRC + GT	0.58	0.66	0.77	0.79	0.79	0.79	0.79	0.69
Set 5: SDS + GT	0.71	0.72	0.78	NS	NS	NS	NS	0.73
Set 6: SIG + GT	0.69	0.74	0.78	NS	NS	NS	NS	0.77
Severe heat stress	Dough development time	Torque	Alveograph W	Alveograph P/L	Alveograph tenacity	Alveograph extensibility	Alveograph P/L*	Loaf volume
Set 1: SRC	NS	0.61	0.74	0.51	0.76	0.54	0.54	0.46
Set 2: SDS	0.68	0.75	0.79	NS	NS	0.66	NS	0.72
Set 3: SIG	0.65	0.75	0.84	NS	0.57	NS	NS	0.62
Set 4: SRC + GT	0.58	0.71	0.82	0.61	0.79	NS	0.69	0.72
Set 5: SDS + GT	0.68	0.75	0.79	0.51	0.57	0.73	0.51	0.77
Set 6: SIG + GT	0.65	0.75	0.84	0.51	0.67	0.65	0.55	0.75

*NS: not significant; the rest of r values were highly significant (p<0.0001); **ALV P/L calculated using predicted alveograph tenacity and alveograph extensibility.

SDSS: SDS sedimentation; SRC: solvent retention capacity; SIG: swelling index of glutenin; GT: grain traits, which could include test weight, thousand kernel weight, grain hardness and grain protein content.