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2 Influence of the final ethanol concentration on the acetification and production rate in the3 wine vinegar process.

4

5 Short title:

- 6 Influence of the final ethanol concentration on the acetification rate.
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8 Authors:

- 9 Silvia Baena-Ruano^a, Carlos Jiménez-Ot^a, Inés M. Santos-Dueñas^a, Jorge E. Jiménez-
- 10 Hornero^b, José L. Bonilla-Venceslada^a, Carmen Álvarez-Cáliz^a, Isidoro García-García^{a*}.

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- 12 Affiliation:
- 13 ^a Departamento de Ingeniería Química, Edificio Marie Curie, Facultad de Ciencias,
- 14 Universidad Córdoba, Campus Universitario de Rabanales, 14071 Córdoba, Spain.
- 15 ^b Departamento de Informática y Análisis Numérico, Edificio Leonardo da Vinci (CTI),
- 16 Escuela Politécnica Superior, Universidad de Córdoba, Campus Universitario de Rabanales,
- 17 14071 Córdoba, Spain.
- 18 * Correspondence to: Isidoro García-García. Phone and Fax: +34 957 218589. E-mail:
- 19 <u>isidoro.garcia@uco.es</u>
- 20

21 Abstract

BACKGROUND: The acetification process still needs an overall study of the variables influencing it in order to establish their optimum values. Based on industrial experience and available literature, including a recently proposed model by the authors, amongst the variables most strongly influencing the acetification process are the ethanol concentration at the time the reactor is unloaded, the unloaded volume and the loading rate. In the scope of ensuring economically efficient industrial production of vinegar, as well as checking the predictions by the aforementioned model, the influence of the final ethanol concentration at unloading time on the mean acetification rate and on productivity has been studied in this work.

5 RESULTS: An increase in the final ethanol concentration from 0.5 to 3.5 % (v/v) increases
6 the mean overall acetification rate and acetic acid production by 38 and 26 %, respectively.

7 The increase is mainly established during the loading phase.

8 **CONCLUSIONS**: The final ethanol concentration is a key variable for the process 9 optimization. If a high rate is desired then a product containing much unused substrate will be 10 obtained, which may be industrially unacceptable. These results suggest the necessity to 11 investigate other possibilities when high values for yield and productivities must to be 12 achieved.

13

Keywords: Vinegar, wine vinegar, *acetobacter*, acetic acid, fed-batch culture, optimization.

16 NOTATION

17	(- <i>r</i> _E)	Ethanol uptake rate, mL ethanol	· (100 mL medium ·	$(\mathbf{h})^{-1} \equiv \% (\mathbf{v}/\mathbf{v}) \cdot \mathbf{h}^{-1}$
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- 18 r_A Acetification rate, g acetic acid \cdot (100 mL medium \cdot h)⁻¹ = % (w/v) \cdot h⁻¹
- 19 P_A Acetic acid production, g acetic acid \cdot h⁻¹
- 20 $(-r_E)_{LP1}$ Mean ethanol uptake rate during loading phase 1, % $(v/v) \cdot h^{-1}$
- 21 $(-r_E)_{LP2}$ Mean ethanol uptake rate during loading phase 2, % (v/v) \cdot h⁻¹
- 22 $(-r_E)_{PP}$ Mean ethanol uptake rate during production phase, $\% (v/v) \cdot h^{-1}$
- 23 $(-r_E)_{Global}$ Mean overall ethanol uptake rate, $\% (v/v) \cdot h^{-1}$

1 INTRODUCTION

Industrially, wine vinegar is obtained mainly by using a semi-continuous process involving submerged acetic acid bacteria¹⁻⁴. Specifically, once the alcohol content falls below a preset level, a portion of the culture medium is unloaded, that remaining in the reactor acting as inoculum for the next load. This allows the production of high-acidity vinegar, facilitates the use of part of the biomass formed in each load to rapidly ferment the next and alters the conditions of the medium in such a way that it can be efficiently used by the most suitable organisms for the intended purpose⁵⁻¹⁰.

9 This operational procedure affords easier control of some variables including the 10 ethanol concentration at unloading time, unloaded volume and loading rate. Such variables 11 influence the concentration and activity of acetic acid bacteria as they act simultaneously on 12 the acidity, ethanol concentration, oxygen supply and even temperature of the medium.

Although a wealth of knowledge currently exists on the vinegar production process,
there remains the need to optimize the previous variables, which may have a significant effect
on the fermentation rate^{4, 11-19}.

Recently, a review with previous attempts for modeling the process as well as a new model proposal has been published^{4, 17,18}. According to this model, the operational conditions depend of the specific type of product to be obtained. If a high rate is desired, the final ethanol content should not be very low, otherwise the bacterial cells may be severely affected by the scarcity of substrate and the high acidity of the medium, which can seriously hinder their reuse on the next load¹⁸.

- This paper reports the results of a study of the influence of the ethanol concentration at unloading time on the productivity and overall rate of the acetification process.
- 24

1 MATERIAL AND METHODS

2 Raw material

3 The raw material used was white wine from the Montilla-Moriles region (Córdoba, 4 Spain) with an ethanol content of 12.0 % $(v/v) \pm 0.3$ and an initial acidity of 0.2 % (w/v). 5 6 **Microorganisms** 7 The inoculum used consisted of a mixed bacterial culture of the genera Acetobacter 8 and Gluconobacter where Acetobacter aceti and Acetobacter pasteurianus were the predominant species²⁰. The inoculum was obtained from a fully operational industrial 9 10 fermentation tank of the firm Grupo SOS, in its Alcolea factory (Córdoba, Spain). 11 12 **Analytical methods** 13 Acidity was determined by acid-base titration and ethanol quantified on-line by means of an Alkosens[®] probe (Heinrich Frings (http://www.frings.com)) 14 15 16 **Fermentation conditions** Tests were conducted in a Frings 8 L fermentation tank, operated in a semi-continuous mode 17 consisting of the following steps: 18 19 Depletion of ethanol in the medium (production phase, PP) to a concentration (1) of 0.5, 1.5, 2.5 or 3.5 % (v/v) at a constant temperature of 31 °C and an also 20 constant air flow-rate of 7.5 L air $h^{-1} \cdot L^{-1}$ medium. Once the desired ethanol 21 22 concentration was reached, 50 % of the tank contents were unloaded. Regarding the loading phase, several strategies are possible. Nevertheless, the 23 (2) aforementioned model by the authors¹⁸, suggest that the charging step must be 24 25 carried out in such a way as to keep the ethanol concentration within the

1 approximate range 5-6 % (v/v). Ethanol levels around and above 6 % (v/v) reduce 2 the proportions of viable cells as well as influence negatively the bacterial activity. So, the tank was slowly loaded (feed rate of $1.2 \text{ L}\cdot\text{h}^{-1}$) to an ethanol concentration 3 never exceeding 5 % (v/v). This was done in two steps: loading phase 1 (LP1) and 4 5 loading phase 2 (LP2). In the first, the tank was loaded in a continuous manner to 6 an ethanol concentration of 5 % (v/v); in the second, more wine was added in a 7 semi-continuous manner until the desired final working volume (8 L) was 8 completed.

9 The bioreactor was fully equipped to operate in an automated mode. Loading, 10 unloading, control and monitoring operations were performed unattended via a previously 11 programmed computer.

Because the primary purpose of this work was to compare the influence of the target operating conditions on the overall rate of the process, the rate had to be previously determined. Provided the total strength remained roughly constant and identical with that of the starting wine throughout the cycle, the ethanol and acetic entrainment losses are negligible, so the mean fermentation rate can be estimated both from the variation of the ethanol concentration during the cycle²¹ and from the final acidity.

The fermentation rate determination from the ethanol concentration, $(-r_E)$, allows one to establish the variation of the acetification rate throughout the cycle, so it is possible to assess the influence of the operational variables on the different steps of the process. Details of how estimating the mean acetification rate via on-line monitored changes in ethanol during a semi-continuous vinegar production cycle can be found elsewhere²¹.

At the same time, the mean acetification rate, r_A , can be easily calculated from the final acidity in the medium at unloading time, the unloaded volume, the cycle duration and the weighted mean of the fermentation broth volume:

$$r_{A} = \frac{\text{final acetic acid concentration } (\% (w/v)) \cdot \text{unloaded volume } (L \text{ medium})}{\text{cycle time } (h) \cdot \text{mean overall volume } (L)} (1)$$

$$gr_{A} = \frac{\text{final acetic acid concentration } (1)}{\text{cycle time } (h) \cdot \text{mean overall volume } (L)}$$

$$gr_{A} = \frac{11.0 \% (w/v) \cdot 4 \text{ L medium}}{31.9 \text{ h} \cdot 7.68 \text{ L}} = 0.18 \frac{\% (w/v)}{\text{h}} (2)$$

9 RESULTS AND DISCUSSION

10 As stated above, wine vinegar is usually obtained by using a semi-continuous 11 fermentation process involving periodic unloading of the fermentation medium. The ethanol 12 concentration present at the time the reactor is unloaded is one of the variables most strongly 13 influencing the overall fermentation rate. In fact, the more markedly ethanol is depleted, the 14 largest is the amount of acid formed -with which bacteria may eventually react. In this work, 15 the influence of ethanol concentrations of 0.5, 1.5, 2.5 and 3.5 % (v/v) were studied. For 16 instant, Figure 1 shows the variation of the ethanol content, acidity and volume of the medium during the acetification cycle at the ethanol concentration of 1.5 % (v/v). The figure, which 17 18 shows the results of 8 tests, clearly exposes the steps involved in the fermentation cycle 19 (particularly the ethanol and volume data). Table 1 list the duration of loading and production 20 phases, the time and final acidity values, as well as the average volume for all the experiments; 21 data are accompanied by their respective standard deviations. Table 2 lists the ethanol uptake 22 rate $(-r_E)$ for each phase and global values, the mean acetification rate r_A values, as well as the 23 production of acetic acid, P_A , in g acetic acid $\cdot h^{-1}$.

Figure 2 shows the acetification rate and acetic acid production percent differences from the lowest levels (viz. those leading to a final ethanol concentration of 0.5 % (v/v)) as well as the mean overall ethanol and acetic acid concentrations for each case. As can be seen, the acetification rate and the acetic acid production increased with increasing ethanol concentration at unloading time by about 38 and 26 %, respectively.

6 Based on the sensitivity of acetic acid bacteria to both the substrate and product, and on 7 changes in the culture medium, one can expect them to perform disparately under different experimental conditions^{4, 22-24}. In fact, our tests exposed differences in mean overall acidity 8 9 and ethanol concentration (Fig. 2), and also in the highest acidity level reached (Table 1). A 10 high acidity is invariably accompanied by a low ethanol concentration. Both can adversely 11 affect the overall rate of the process, ethanol because it is the limiting substrate and acetic acid 12 because of its inhibitory effect increases with increasing concentration⁴. In this case, the 13 known negative influence that high ethanol concentrations can have on cell viability demonstrated by authors elsewere⁴, it is not a problem because of the followed loading 14 15 strategy by which the ethanol concentration was never higher than 5 % (v/v). Nevertheless, the 16 acetic acid concentration could be the key factor for explaining the differences observed in this 17 work. Indeed, a decrease in bacterial viability is normally observed as the mean acetic acid concentration increases and, specially, when maxima (final) acidities reach $11 \% (w/v)^4$. 18

The process can also be affected by changes occurring between unloading and the end of the loading process; such changes become more marked as the substrate is depleted as well as the loading step is shortened. For instant, Figure 3 shows regression for the experimental variation of ethanol content and volume of the medium during the cycle at each studied final ethanol concentration. From the first, it can be seen that, during the loading phase, bacteria are subjected to important differences in the ethanol concentration (and therefore in acidity) which may have a negative influence in the process. On the other side, from the second, it can be

seen that, from a kinetic point of view, main differences are found in the loading phase 2.
 During this step, where the ethanol concentration is kept constant, the fermentation rate can be
 estimated from the temporal variation of the volume²¹.

4

Table 2 lists the ethanol uptake rates for each phase throughout the cycle.

5 Provide ethanol uptake rates during loading phase 1 (Table 2) have not statistically 6 significant differences (one way ANOVA test), must be in loading phase 2 and production 7 phase where the increase of acetification rate is established. As can be calculated from data 8 listed on Table 2, the uptake ethanol rate for loading phase 2 and production phase increased 9 with final ethanol concentration by about 82 and 13 %, respectively. Nevertheless the overall 10 rate of ethanol uptake increased just by about 40 % since a weighted average as a function of 11 the proportion of time taken by each phase has to be considered. In any case, it is clear that 12 main differences are found in loading phase 2.

13 Based on the previous results, which contribute to validate the previously proposed model by the authors¹⁸, obtaining a high rate for the process entails ensuring a high ethanol 14 15 concentration at unloading time; this, however, considerably reduces the acetification yield 16 through the presence of a substantial amount of ethanol in the unloaded liquid. This 17 shortcoming can be circumvented, by using two serially arranged reactors. The two reactors 18 can be optimized in such a way as to ensure that the first is unloaded at a high concentration of 19 ethanol and the second depletes it before it receives a new load from the first; this study is at 20 present going on.

21

22 **5.** Conclusions

The semi-continuous wine acetification process usually employed by the vinegar production industry can be improved as regards overall rate by optimizing some easily adjusted process variables including the ethanol concentration at unloading time, unloaded 8

1 volume and loading charge. In this work, the influence of the ethanol concentration on the 2 production and mean acetification rate was studied. Based on the results, the acetification rate 3 and the production of acetic acid increase substantially with increase in the ethanol 4 concentration. Also it is concluded that the increase of the acetification rate is established 5 mainly during the loading phase. However, a product containing much unused substrate is 6 industrially unacceptable. For this reason, two serially arranged reactors, usually available in 7 industry, must be optimized in such a way that the first one must ensure a high acetification 8 rate, but also a final ethanol concentration that can be easily depleted in the second reactor 9 before it receives a new load from the first. This procedure should give mean overall 10 acetification rates higher than those typically obtained when the substrate is depleted much 11 more in the first reactor.

12

13 Acknowledgements

The authors wish to acknowledge funding of this research (AGL2002-01712,
PET2006-0827) by Spain's Ministry of Science and Technology (MCyT) and Ministry of
Science and Education. Also, they wish to thank the firm Grupo SOS (Spain) for valuable help
and advice.

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19 REFERENCES

20

1 De Ory I, Romero LE and Cantero D, Maximun yield acetic acid fermenter. Comparative
fed-batch and continuous operation studies at pilot plant scales, *Bioprocess Eng* 21: 187–190
(1999).

24

25 2 Ebner H, Sellmer S and Follmann H, Vinegar, in Ullmann's Encyclopedia of Industrial
 9

1	Chemistry VCH, ed by Elvers B and Hawkins S. VCH, Weinheim, 27: 403–418 (1996).
2	
3	3 Hromatka O and Ebner H, Investigations of the vinegar fermentation III. The influence of
4	aeration on submerged fermentation. Enzymologia 15 (2): 57-69 (1951).
5	
6	4 Jiménez-Hornero JE, Santos-Dueñas IM and García-García I, Optimization of
7	biotechnological processes. The acetic acid fermentation. Part I: The proposed model,
8	<i>Biochem Eng</i> J 45 : 1-6 (2009).
9	
10	5 Arnold S, Becker T, Delgado A, Emde F and Enenkel A, Optimizing high strength acetic
11	acid bioprocess by cognitive methods in an unsteady state cultivation. J Biotechnol 97: 133-
12	145 (2002).
13	
14	6 Ebner H, Sellmer S and Follmann H, Acetic Acid, in <i>Biotechnology</i> , ed by Rehm HJ and
15	Reed G. VCH, Weinheim, 6: 381-401 (1996).
16	
17	7 Levonen E and Llaguno C, Tecnología de la fabricación de vinagre. Revista Agroquímica de
18	Tecnología Alimentaria 18: 289–296 (1983).
19	
20	8 Llaguno C, Marchena M and Polo C, El vinagre de vino. CSIC. Madrid, Spain (1991).
21	
22	9 Suárez JA and Iñigo B, Microbiología Enológica. Fundamentos de vinificación. Ediciones
23	Mundi-Prensa. Madrid, Spain (1992).
24	

1	10 Tesfaye W, Morales ML, García-Parrilla MC and Troncoso AM, Wine vinegar:
2	technology, authenticity and quality evaluation, Trends Food Sci Technol 13: 12-21 (2002).
3	
4	11 Garrido-Vidal D, Pizarro C and González-Sáiz JM, Study of process variables in industrial
5	acetic fermentation by a continuous pilot fermentor and response surfaces. Biotechnol Prog
6	19 : 1468–1479 (2003).
7	
8	12 Gómez JM and Cantero D, Kinetics of substrate consumption and product formation in
9	closed acetic fermentation systems. Bioprocess Eng 18: 439-444 (1998).
10	
11	13 González-Sáiz JM, Pizarro C and Garrido-Vidal D, Evaluation of kinetic models for
12	industrial acetic fermentation: proposal of a new model optimized by genetic algorithms.
13	Biotechno. Prog 19(2): 599-611 (2003).
14	
15	14 Romero LE, Gómez JM, Caro I and Cantero D, A kinetic model for growth of Acetobacter
16	aceti in sumerged culture. Chem Eng J and Biochem Eng J 54: B15-B24 (1994).
17	
18	15 Macías M, Caro I and Cantero D. Optimum operating conditions in closed-system
19	industrial acetifiers (batch operation): a study by computer simulation. Chem Eng J and
20	<i>Biochem Eng J</i> 62 : 183-191 (1996).
21	
22	16 Macías M, Caro I and Cantero D, Optimum operating conditions in closed-system
23	industrial acetifiers (semi-continuous operation): a study by computer simulation. Chem Eng J
24	65 : 201-207 (1997).
25	

1	17 Jiménez-Hornero JE, Santos-Dueñas IM and García-García I, Optimization of
2	biotechnological processes. The acetic acid fermentation. Part II: Practical identifiability
3	analysis and parameter estimation. Biochem Eng J 45: 7-21 (2009).
4	
5	18 Jiménez-Hornero JE, Santos-Dueñas IM and García-García I Optimization of
6	biotechnological processes. The acetic acid fermentation. Part III: Dynamic optimization.
7	Biochem Eng J 45 : 22-29 (2009).
8	
9	19 Nishiwaki A and Dunn IJ, Analysis of acetic acid productivity in a continuous two-stage
10	bioreactor with cell recycling. J Chem Technol Biotechnol 80: 371-375 (2005).
11	
12	20 Barja F, González A, Mesa-Díaz MM, Macías M and Cantero D, Molecular and
13	morphological characterization of acetic acid bacteria from industrial fermenters of wine
14	vinegar production. In: Book of Abstracts, International Symposium of Vinegars and Acetic
15	Acid Bacteria, ed by Giudici P, Reggio Emilia, Italia, 68 (2005).
16	
17	21 García-García I, Cantero-Moreno D, Jiménez-Ot C, Baena-Ruano S, Jiménez-Hornero JE,
18	Santos-Dueñas IM, Bonilla-Venceslada JL and Barja F, Estimating the mean acetification rate
19	via on-line monitored changes in ethanol during a semi-continuous vinegar production cycle. J
20	<i>Food Eng</i> 80 : 460-464 (2007).
21	
22	22 Nickol GB, Vinegar, in Microbiology technology. 2nd ed by Peppler H J and Perlman D
23	New York, Academic Press 155–172 (1979).
24	
25	23 Lu SF, Lee FL and Chen HK, A thermotolerant and high acetic acid-producing bacterium

- 1 Acetobacter sp. J Appl Microbiol **86**: 55–62 (1999).
- 2
- 3 24 Lasko DR, Zamboni N and Sauer U, Bacterial response to acetate challenge: a comparison
- 4 of tolerance among species. *Appl Microbiol Biotechnol* **54**: 243–247 (2000).
- 5

2 Table 1

Final ethanol concentration, %	0.5	1.5	2.5	3.5
(v/v)				
Number of cycles	6	8	10	29
Duration of loading phase 1, h	2.5 ± 0.1	2.1 ± 0.1	1.4 ± 0.1	0.9 ± 0.1
Duration of loading phase 2, h	6.8 ± 0.4	8.3 ± 0.7	8.9 ± 0.6	11.0 ± 0.8
Duration of production phase, h	22.6 ± 0.9	16.9 ± 0.8	12.0 ± 0.9	6.8 ± 1.2
Cycle duration, h	31.9 ± 0.8	27.3 ± 0.4	22.3 ± 0.5	18.7 ± 0.9
Final acidity (as acetic acid), %	11.0 ± 0.2	10.1 ± 0.2	9.2 ± 0.15	8.1 ± 0.1
(w/v)				
Mean total volume, L	7.68 ± 0.06	7.54 ± 0.03	7.37 ± 0.02	7.00 ± 0.02

2 Table 2

Final ethanol concentration, %	0.5	1.5	2.5	3.5
(v/v)				
$(-r_E)_{LP1}, \% (v/v) \cdot h^{-1}$	0.13 ± 0.10	0.23 ± 0.24	0.20 ± 0.17	0.29 ± 0.29
$(-r_E)_{LP2}, \% (v/v) \cdot h^{-1}$	0.15 ± 0.01	0.19 ± 0.01	0.25 ± 0.01	0.28 ± 0.01
$(-r_E)_{PP}, \% (v/v) \cdot h^{-1}$	0.20 ± 0.01	0.21 ± 0.01	0.22 ± 0.01	0.23 ± 0.01
$(-r_E)_{Global}, \% (v/v) \cdot h^{-1}$	0.19 ± 0.01	0.20 ± 0.02	0.23 ± 0.02	0.26 ± 0.03
r_A , % (w/v) · h ⁻¹	0.18 ± 0.01	$0.\overline{20\pm0.01}$	0.22 ± 0.01	0.25 ± 0.01
P_A , g acetic acid \cdot h ⁻¹	13.8 ± 0.4	14.8 ± 0.4	$\overline{16.5\pm0.5}$	17.4 ± 0.9

- All figures were created by SigmaPlot for Windows Version 11.0
- Figure 1



Figure 2



1 Figure 3



1 Lege	ends
--------	------

3 Table 1

4 Experimental phase and cycle duration, final acidity and mean overall volume obtained under

- 5 different experimental conditions plus their standard deviations.
- 6

7 Table 2

8 Phase and global ethanol uptake rate, mean acetification rate and acetic acid production,

9 accompanied by their standard deviations.

10

11 Figure 1

12 Variation of the ethanol concentration, volume and acidity of the medium during the

13 fermentation cycle. Final ethanol concentration at unloading time: 1.5 % (v/v). (LP1: loading

14 phase 1; LP2: loading phase 2; PP: production phase).

15

16 Figure 2

17 Acetification rate and acetic acid production percent differences from the lowest levels as well

18 as the mean overall ethanol and acetic acid concentrations. Bars represent standard deviations.

19

20 Figure 3

21 Regression for the experimental variation of ethanol content and volume of the medium during

22 the cycle at each studied final ethanol concentration.

23