



# Article Arbosana Olive Is Self-Incompatible, but Inter-Compatible with Some Other Low-Vigor Olive Cultivars

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**Abstract:** Trendy high-density olive fields are often monovarietal orchards, mostly using the cultivar Arbequina. However, Arbequina shows a strong self-incompatibility response, and its yields depend on wind cross-pollination, which is not always available. With the aim of finding suitable self-compatible cultivars that can replace Arbequina, we evaluated pollen–pistil interaction, fruit set and seed paternity in Arbosana under different pollination treatments: self-pollination, open-pollination and three cross-pollination treatments: × Arbequina, × Sikitita and × Koroneiki. All these cultivars are low-vigor cultivars suitable for high-density orchards, making them potential pollinizers for Arbosana. The results show that Arbosana is also self-incompatible with a strong reduction in fruit set due to a lower fertilization level caused by a strong inhibition of pollen tube growth in self-pollinated flowers. Seed-paternity analyses confirmed the self-pollination were, in fact, a product of cross-fertilization. In conclusion, we recommend against the use of Arbosana in large monovarietal orchards. On the contrary, good results were obtained under cross-pollination with Sikitia, Arbequina and Koroneiki pollen, allowing us to recommend them as pollinizers for Arbosana in appropriate pollination designs. This is the first time Arbosana self-incompatibility has been reported.

**Keywords:** *Olea europaea*; self-incompatibility; high-density orchards; pollen–pistil interactions; fruit set; seed paternity

# 1. Introduction

World olive (*Olea europaea* L.) oil production was just over 3.3 million tons, and nearly 23.5 million tons of olives were produced on 10.3 million hectares in 2021, which is the last year from which we have final data [1]. In that year, Spain, the world's largest producer of olive oil, produced 1.5 million tons of olive oil (44.6% of the total) and 8.3 million tons of olives (35.1%) on 2.6 million hectares [1]. However, olive tree cultivation is facing a dramatic change worldwide. After centuries of rain-fed, low-density orchards planted with local cultivars, olive growing is rapidly transitioning in the last few decades to large, super high-density irrigated orchards [2,3], which are mostly achieved using one single cultivar: Arbequina [4]. However, Arbequina is self-incompatible [5], and the high yields needed to control vegetative growth strongly depend on cross-pollination from neighboring cultivars that act as unsuspected pollinizers [6,7]. This benefit is possible in traditional olive countries due to the airborne condition and massive production of pollen grains in this wind-pollinated taxon. In fact, several authors have demonstrated through seed



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**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). paternity analyses that most fruit produced in supposedly monovarietal orchards are sired by cross-pollination from distant sources [8,9]. Unfortunately, this free cross-pollination does not exist in new, recent areas of olive cultivation, and the varietal mosaic in traditional olive countries is becoming more and more reduced, compromising the benefits of this free open-pollination.

Olive blooms in panicles developed in 1-year-old shoots. The panicle is the unit of fructification in olive and bears staminate (pistil-aborted) as well as hermaphrodite flowers in a variable number (usually between 12 and more than 20). The hermaphrodite flower is composed of four greenish small sepals, four larger white petals, two opposite stamens producing plenty of pollen grains and a pistil where a large stigma stands out. The style of the pistil is, on the contrary, short. The ovary contains four ovules in two locules, although usually only one ovule is fertilized and becomes the seed of the fruit (drupe). Staminate flowers, produced in a variable proportion in the panicle of the olive, share the basic morphology of the hermaphrodite flowers, but the pistil in them is rudimentary or completely absent. Olive flowers are receptive from anthesis, when anthers dehiscence starts and the stigma receptivity allows pollen adhesion. Olive is considered preferentially allogamous for most authors with a preference for cross-pollen to achieve the fertilization of the ovule becoming seed. The preferential allogamy is based on a self-incompatibility system that is still under dispute, although recent work strongly suggests that, in olive, self-incompatibility corresponds to the sporophytic system [10,11].

Given the preferential allogamy of most olive cultivars, Arbequina included, and the increasing interest in very large monovarietal high-density orchards, we have studied in two consecutive years the self-incompatibility behavior of Arbosana, which is a low-vigor cultivar often selected as an alternative to Arbequina in different countries and growing conditions [12–15]. Despite its wide use in high-density orchards, Arbosana self-incompatibility response has not yet been determined.

#### 2. Materials and Methods

## 2.1. Plant Material and Study Site

An experiment to determine Arbosana self-incompatibility response was carried out during the seasons 2022 and 2023 on adult olive trees planted in 2011 in a high-density orchard located on the Rabanales Campus of the University of Córdoba (Córdoba, Spain; 37°56′05″ N, 4°43′00″ W, at 160 m altitude). The trees are drip-irrigated with volumes between 1000 and 2000 m<sup>3</sup>/ha, and pests and diseases are controlled by IPM programs. Harvest and pruning is performed mechanically, as usual in high-density orchards trained in hedgerow systems. Arbosana is noted for its reduced vigor, good adaptation to super high-density plantations, early production and heavy and regular yields of very good quality olive oil. Its good agronomic performance justifies its increased acreage. Arbosana accounts for around 25% of high-density orchards and represents a clear alternative to the main cultivar Arbequina for warm areas, such as the experimental site, where it produces as much as Arbequina.

The average temperature at the experimental site during the bloom and fruit set periods (April and May) of 2022 was 18.8 °C with an average relative humidity of 60.3% and total rainfall reaching a significant volume of 78.7 mm in less than two months. The 2023 season was slightly warmer (an average of 20.5 °C) and drier (average relative humidity of 48.1%) with less rainfall totaling 32.6 mm. The weather conditions during the bloom and fruit set periods in 2022 and 2023 are shown in Figure 1. Climactic data were retrieved from a weather station located nearby at the Campus of the University of Córdoba.

In the experiments, four adult trees of Arbosana were used as replications. The trees were  $4 \times 2$  m spaced and trained as a hedgerow in a multivarietal orchard. Adjacent tree rows of the cultivars Arbequina, Sikitita, Koroneiki and Tosca were included in the orchard. The orchard layout showing the tree rows arrangement of the different cultivars is depicted in Figure 2.



**Figure 1.** Temperature, humidity and rainfall during the experimental seasons 2022 (**a**) and 2023 (**b**). Data retrieved from a weather station sited nearby at the Campus of the University of Córdoba.



**Figure 2.** Experimental orchard layout. In blue, tree row of Arbosana (AS), in green Arbequina (AQ) trees, in pink Tosca (TO), in yellow Sikitita (SK) and in red Koroneiki (KO) rows.

Arbequina is, so far, the most popular olive cultivar for high-density orchards based on its high and regular productivity, precocious yield and good olive oil quality. Arbequina accounts for around 65% of the area of the high-density plantations carried out all around the world (last estimations are near 100,000 hectares). Sikitita, on the contrary, is a new olive cultivar recently released from the UCO olive-breeding program, suitable for highdensity orchards due to its very much reduced plant vigor. It presents the lowest alternate bearing index of the cultivars constituting the experimental orchard. Koroneiki is a Greek cultivar with excellent olive oil quality that is well adapted to semi-arid zones where is very productive, but it shows a higher tendency to alternate bearing. Koroneiki also exhibits higher vigor than the other cultivars tested. Finally, Tosca is the main Italian cultivar used for high-density orchards. It is also a low-vigor cultivar, maturing early in the season, allowing, therefore, harvesting before some other cultivars. In comparative trials carried out from 2013 to 2018 in the same experimental orchard, Tosca has shown lower productivity than Arbequina, Arbosana, Sikitita and Koroneiki [16].

Four experimental trees of 'Arbosana' were chosen for their uniform and high level of flowering. In 2022, each of these trees received four pollination treatments: self-pollination, open pollination, cross-pollination with Arbequina pollen, and cross-pollination with Sikitita pollen. In the 2023 season, we added a new cross-pollination treatment using Koroneiki pollen.

The self-pollination treatment was achieved by bagging, before bloom, 1-year-old flowering shoots with hand-made tissue paper bags. These hand-made paper bags, although fragile in the event of rain, do not substantially modify the temperature and humidity conditions inside them. Cross-pollination was performed by applying fresh pollen collected from the selected pollen donor cultivars to the stigmas of the open flowers of Arbosana using a fine camel paintbrush. Cross-pollen was repeatedly applied to the open flowers every other day. After performing hand pollination, cross-pollinated shoots were re-bagged to avoid pollen entrance from other neighboring cultivars. Pollen grains used for pollination were repeatedly harvested from the neighboring trees on the same day of pollination. Pollinations were performed between 25 April and 1 May in 2022 and from 13 April to 22 April in 2023. Emasculation of the flowers was not performed, so these flowers received their own pollen plus the pollen from the selected pollinizer. Open-pollinated flowers were left unbagged. Open pollination in a multivarietal orchard represents the optimal pollination treatment due to the unrestricted and continuous pollen arrival from different sources and different pollen donor trees.

#### 2.2. Initial and Final Fruit Measurements

To determine Arbosana response to these pollination treatments, we measured initial and final fruit set for each pollination treatment in four trees as replications. Fruit set was calculated as the number of fruit per inflorescence measured in ten shoots per tree in 2022 and in eight shoots per tree in 2023. The selected shoots bore ten inflorescences each. That uniform flowering level was achieved by thinning the excess inflorescences when necessary. The shoots were located all around the trees at observer height. The shoots were bagged (except open-pollination treatment) before bloom and received the corresponding pollination treatment every other day and at least three times as bloom progressed. After performing hand cross-pollination, the shoots were bagged again. Initial fruit set was calculated as the number of enlarged ovaries (fruitlets) per inflorescence two weeks after bloom. Final fruit set was estimated as the number of fruitlets per inflorescence seven weeks after bloom. With final fruit set data, we calculated the Index of Self-Incompatibility (ISI) by dividing self-pollination fruit set by open- (and cross-) pollination fruit set [17].

## 2.3. Pollen–Pistil Interaction

For the study of the pollen–pistil interaction, samples of 20 flowers (occasionally less) of each pollination treatment were collected 1, 2, 4 and 8 days after pollination (dap) in 2022, and 2, 4 and 8 dap in 2023. Pollination of these flowers was carried out the day of anthesis and one day after. Anthesis date was ensured by removing the open flowers on a given day and all closed flowers the next day, so all remaining flowers opened between one given day and the next. Pollination treatments were applied as explained before for fruit set measurements. The sampled flowers were immediately fixed in a FAE solution made with formalin, glacial acetic acid and 70% ethanol in a 1:2:17 ratio by volume and sent to the lab. These samples were kept in vials in the fridge at 4 °C until the evaluation of pollen–pistil interaction under fluorescence microscopy. The parameters analyzed were pollen adhesion, pollen germination, pollen tube growth and fertilization. For processing the flowers, they were softened during 8 h with a solution of NaOH 1N, washed in running water, stained with aniline blue, and gently squashed before observation under fluorescence microscopy using a Labophot microscope (Nikon, Tokyo, Japan).

Pollen adhesion and germination was measured soon after anthesis (1, 2 and 4 dap in 2022, and 2 and 4 dap in 2023). Pollen adhesion was calculated by counting the number of pollen grains adhered to the stigma of the flowers. Pollen germination was estimated as the number of them showing a pollen tube of at least the diameter of the pollen grain. Pollen tube growth and fertilization were evaluated 2, 4 and 8 dap. Pollen tube growth was estimated on a scale from 0 to 3 as follows: 0 for no pollen tubes growing in the style, a value of 1 when 1 to 5 pollen tubes were present in the upper part of the style, a value of 2 when 5 to 25 pollen tubes were present in the style, and a value of 3 for more than 25 pollen tubes per flower growing toward the ovary. For the evaluation of fertilization, the four ovules of each flower were dissected from the ovary and observed under fluorescence microscopy. The presence of a pollen tube in the micropyle of the ovules, usually in only

one, was considered proof of fertilization. This procedure was followed the same way in both seasons.

## 2.4. Seed-Paternity Analyses

Finally, to determine the pollen genotype achieving ovule fertilization, samples of fruit from the shoots used for fruit set analyses were randomly selected (when in excess) and collected soon before harvest to determine the paternity of their seeds. Based on the 2022 results, we decided to limit the analyses of seed paternity in 2023 samples to open-, selfand to the new cross-pollination treatment using Koroneiki pollen. Seed-paternity analyses were carried out following the procedure described by Seifi et al. [18], slightly modified. In addition to the genotyping of the seed embryos, fresh olive leaves were sampled from the five different genotypes (Tosca, Sikitita, Koroneiki, Arbequina and Arbosana) present in the experimental orchard and acting as potential pollen donors. Genomic DNA was extracted from the embryos and leaves using a NucleoSpin<sup>™</sup> Plant II kit (Macherey-Nagel, Hoerdt, France) according to the manufacturer's instructions. Eight polymorphic SSR markers were used for genotyping: ssrOeUA-DCA-(3, 5, 15, 18) [19], GAPU-(71B, 101) [20], and EMO-(3, 90) [21]. PCR reaction was performed in a final volume of 12.5  $\mu$ L containing 1x supplied AllTaq PCR buffer, 1x supplied Q-Solution, 2 mM MgCl2, dNTP mix (0.2 mM of each dNTP), 1.25 U of AllTaq DNA polymerase (Qiagen, Hilden, Germany), 0.2 mM reverse locus specific primer, M13(-21) tail labeled with fluorescent dye 6-FAM, VIC, NED or PET (Applied Biosystems, Foster City, CA, USA), 0.1 mM elongated forward primer, and 4  $\mu$ L of undiluted template DNA for olive embryos or 40 ng of template DNA for potential pollen donors. The amplification was performed on a SimpliAmp<sup>™</sup> Thermal Cycler (Thermo Fisher Scientific, Waltham, MA, USA) according to the two-step touchdown amplification profile [22]. One  $\mu$ L of PCR product mix, including amplicons with four different fluorescent dyes, was added to 10.7 µL Hi-Di<sup>TM</sup> Formamide (Thermo Fisher Scientific, Waltham, MA, USA) and GeneScan<sup>™</sup> 500 LIZ size standard (Thermo Fisher Scientific, Waltham, MA, USA) and analyzed on a SeqStudio<sup>TM</sup> Genetic Analyzer (Thermo Fisher Scientific, Waltham, MA, USA). Electropherograms were scored with GeneMapper version 5 software (Thermo Fisher Scientific, Waltham, MA, USA).

In the first step of the analysis, the five potential pollen donor cultivars cited above were genotyped to obtain their reference profiles and then all embryos were genotyped. In a second step, paternity analysis was performed for all embryos produced from open pollination and for those from self-pollination and cross-pollination treatments in which the presumed father alleles were not detected. Then, the potential pollen donors were identified using paternity analyses using CERVUS 3.0 software (http://www.fieldgenetics. com; accessed on 20 June 2024) [23]. Each CERVUS run consisted of an allele frequency analysis followed by a simulation in which the number of potential fathers was set to five (the cultivars equally represented in the orchard), while the proportion of candidate fathers sampled was set to 80%. The proportion of typed loci was set to the percentage estimated by the allele frequency analysis. A minimum of four typed loci were required for the progeny to be analyzed for paternity, and the number of simulated progenies was set to 100,000. The genotyping error in the simulation and in the assignment of the most likely pollen donor was set at 1%, as possible errors can occur during the allele-calling phase. Finally, the genotypes of the embryos and the available genotypes for the potential pollen donor cultivars were used to calculate the natural logarithm of the likelihood ratio (LOD) scores. These ratios compare the probability that an individual is the parent of a given progeny divided by the probability that these two individuals are unrelated. The genotype with the highest LOD score above the threshold (determined based on the simulation) for a given parent-offspring pair is considered the most likely pollen donor. If the pollen parent was not among the cultivars present in the experimental orchard, it was assumed that the pollen originated from olive cultivars grown outside the orchard.

## 2.5. Statistical Analyses

Fruit set data were analyzed by analysis of variance and means were compared by Tukey's test (p < 0.05) using Statistix 8.0 (Analytical Software, Tallahassee, FL, USA). Fertilization levels in the different treatments were compared by Chi-square tests using the same software.

## 3. Results

# 3.1. Initial and Final Fruit Set

Arbosana behaved as a self-incompatible cultivar. In this regard, self-pollination resulted in a significant and important reduction in the final fruit set compared to open pollination, as well as compared to cross-pollination with Arbequina pollen, with Sikitita pollen and, in 2023, with Koroneiki pollen too (Table 1). The results were similar in 2022 and 2023, although in the second season, fruit set under self-pollination was slightly higher. There were no significant differences between open-pollination treatment and cross-pollination in terms of final fruit set, indicating that all assessed pollinizers may equally serve as effective pollen donors for Arbosana. However, in 2022, significant differences were found between open- versus cross-pollination treatments in the initial fruit set, being higher in the former.

Table 1. Arbosana fruit set in response to different pollination treatments. Seasons 2022 and 2023.

Pollination Treatments	2022 Season			2023 Season			
	Initial Fruit Set *	Final Fruit Set	ISI **	Initial Fruit Set	Final Fruit Set	ISI	
Self-pollination	0.23 c ***	0.18 b		0.43 b	0.32 b		
Open-pollination	3.08 a	1.23 a	0.15	2.32 a	1.23 a	0.26	
$\times$ Arbequina	1.73 b	0.85 a	0.21	2.17 a	1.06 a	0.30	
× Sikitita	1.78 b	1.00 a	0.18	2.37 a	1.25 a	0.26	
imes Koroneiki	_	-		2.53 a	1.13 a	0.28	
p	< 0.0001	< 0.0001		0.0012	0.0002		

\* Fruit set = fruit/inflorescence 18 and 48 days after bloom measured in 8–10 shoots per replication (tree) each bearing 10 inflorescences (panicles). \*\* ISI = final fruit set under self-pollination/final fruit set under openor cross-pollination according to Zapata and Arroyo [17]. \*\*\* Mean comparison in columns by Tukey's test at p < 0.05. For each column, different letters indicate a statistical significance.

Occasionally, a disparity of crop load in branches under self-pollination was observed, with some bagged shoots bearing no or very few fruit, while others produced as many fruit as cross-pollination treatments. We did not exclude the data from those shoots in fruit set calculations, although they slightly increased the average fruit set for self-pollination (from hypothetical 0.07 fruit per inflorescence to 0.18 in 2022, and from 0.19 to 0.32 in 2023). The Index of Self-Incompatibility (ISI) in 2022 ranged between 0.15, when self-pollination is compared to open pollination, to 0.21, when Arbequina pollen was applied. In 2023, ISI fluctuated between 0.26 for open pollination and cross-pollination with Sikitita pollen and 0.30 when using again Arbequina as the pollen donor cultivar.

## 3.2. Pollen-Pistil Interaction

Pollen adhesion was generally high, especially in 2023 (Table 2). In 2022, self-pollinated flowers had an average of 155 pollen grains adhered to the stigma, while open pollination had 205 pollen grains per flower and substantially more under cross-pollination treatments (Table 2). Pollen germination in percentage was, on the contrary, a little bit higher for self- and open pollination, with an average of 26.5 and 37.1%, respectively, while it was 24.2% for Arbequina pollen and 23.6% for Sikitita pollen. A similar trend was observed in 2023 with lower pollen adhesion for self-pollination, and low too for cross-pollinated flowers using Koroneiki pollen. On the contrary, higher pollen adhesion was observed for open-pollination and for other cross-pollination treatments, particularly with Sikitita pollen (33% more pollen adhered per stigma for open and cross-pollination with Arbequina and 64% more when using Sikitita pollen). The percentage of pollen germination was

lower in this season (between 8.6 and 13.7%, depending on the treatment), especially in the treatments with higher pollen adhesion. Nonetheless, on average, the flowers had between 31 and 63 pollen grains germinated with only a few flowers completely lacking pollen grains (one in self-pollination, and one in cross-pollination with Koroneiki pollen) (Table 2).

**Table 2.** Pollen adhesion and germination in Arbosana flowers in response to different pollination treatments. Average  $\pm$  SE of values measured 1, 2 and 4 days after pollination (dap) in 2022 season, and 2 and 4 dap in 2023 season.

	2022	Season	2023 Season		
Pollination Treatments	Pollen Adhesion * Pollen Germination		Pollen Adhesion	Pollen Germination	
Self-pollination	$155\pm32$	$41 \pm 11$	$267\pm53$	$31\pm5$	
Open-pollination	$205\pm32$	$76\pm14$	$402\pm82$	$50\pm9$	
× Arbequina	$339\pm46$	$82 \pm 13$	$397\pm62$	$51\pm 6$	
× Sikitita	$310\pm 61$	$73\pm18$	$732\pm42$	$63\pm8$	
imes Koroneiki	-	-	$227\pm42$	$31\pm4$	

\* Pollen grains adhered per stigma. \*\* Pollen grains germinated on the surface of the stigma.

The most contrasting parameter between self- versus open- and cross-pollination treatments was the growth of the pollen tube in the stigma and style of the flowers. While in open and cross-pollination it was common to observe more than 25 pollen tubes growing in the style (Figure 3b), in self-pollinated flowers, it was rare to see pollen tubes growing beyond the first layers of cells of the stigma, and when some pollen tubes did form in self-pollinated flowers, they crossed the style in a low number (Figure 3a).



**Figure 3.** Representative pictures of pollen tube growth in self-pollination (**a**) and in cross-pollination treatments (**b**) in Arbosana flowers squashed, stained using aniline blue and observed under fluorescence microscopy. Arrow marks pollen tube in self-pollinated flower.

The clear inhibition of pollen tube growth in the stigma and style of self-pollinated flowers was quantified using a scale ranging from 0 (no pollen tubes in the style) to 3 (more than 25 pollen tubes per flower in the style). Thus, the average value of the scale for pollen tubes growing in the style shows the differences among treatments in this parameter (Table 3). In this regard, self-pollinated flowers averaged a value of 0.38 and 0.84 in 2022 and 2023, respectively, with over 89% of the flowers showing no, or fewer than five, pollen tubes in the style. In contrast, open pollination achieved mean values of 1.87 and 1.91 for

the same years, with 65 to 75% of flowers exhibiting massive pollen tube growth. These latter values were very similar to those observed in cross-pollination with Sikitita pollen (1.90 and 1.85), where 70 to 74% of flowers showed a high number of pollen tubes growing in the style in 2022 and 2023, respectively. The most effective treatment in terms of pollen tube growth was cross-pollination with Arbequina pollen, averaging values of 2.11 and 2.48 in 2022 and 2023, respectively, with 62 and 88% of flowers displaying robust pollen tube growth. Koroneiki pollen also promoted massive pollen tube growth in Arbosana flowers, averaging a scale value of 2.23 with 85% of flowers in the rank of 2 or 3 in the scale in 2023 (the only year in which this treatment performed).

**Table 3.** Pollen tube growth and fertilization in Arbosana flowers in response to different pollination treatments. Average of measurements performed 2, 4 and 8 days after pollination. Seasons 2022 and 2023.

	2022 Sea	ason	2023 Season		
Pollination Treatments	Pollen Tube Growth *	Fertilization **	Pollen Tube Growth *	Fertilization **	
Self-pollination	0.38	6.7 b ***	0.84	15.5 c	
Open-pollination	1.87	27.1 a	1.91	62.5 a	
$\times$ Arbequina	2.11	25.0 a	2.48	25.0 bc	
× Sikitita	1.90	20.7 a	1.85	28.6 bc	
imes Koroneiki	_	-	2.23	41.1 b	

\* Pollen tube growth measured in a scale from 0 (absence of pollen tubes growing in the upper part of the style) and 3 (more than 25 pollen tube present in the upper part of the style). \*\* Percentage of flowers with a pollen tube in the micropyle of one of the four ovules. \*\*\* Fertilization original values compared by Chi-square tests. For each column, different letters indicate a statistical significance at p < 0.05.

Due to the inhibition of pollen tube growth under self-pollination, fertilization rates were significantly reduced. Fertilization levels in this treatment averaged 6.7% of the flowers in 2022 and 15.5% in 2023, which were significantly lower than the average levels of fertilization obtained under open pollination in both years (27.1% in 2022 and 62.5% in 2023) (Table 3). Both cross-pollination treatments also significantly enhanced fertilization levels in 2022 with lower increases observed in 2023. Only open and cross-pollination with Koroneiki produced significantly higher values in 2023. Unusually high levels of self-fertilization found 2 dap led to non-statistically significant differences compared to Arbequina and Sikitita in this second season.

#### 3.3. Seed-Paternity Analyses

In the 2022 season, 222 embryos were processed: 65 from open pollination, 57 from crosses with Arbequina pollen, 75 from crosses with Sikitita pollen, and 25 from self-pollination, as fruit from the latter were less abundant. Out of these 222 embryos, clear seed paternity was established for 188 samples through the LOD scores calculations (Table 4). The remaining 34 embryos had LOD scores lower than the established threshold (0.93) and are, therefore, not considered in the subsequent Discussion, but they are presented in Supplementary Table S1. In the 2023 season, we analyzed 116 embryos: 76 from seeds of open pollination, 20 from seeds of self-pollination, and 20 from cross-pollination treatment with Koroneiki pollen. Out of these 116 seeds, clear paternity was determined for 91 embryos (61 embryos from open pollination, and 18 and 12 for self- and cross-pollination, respectively; Table 5). The more likely genotypes fathering the remaining embryos, which had LOD values below the threshold (0.88), are shown in Supplementary Table S2. These data are not further considered for the Discussion.

Seed-paternity analyses revealed the existence of pollen contamination inside the paper bags, which was particularly noticeable under self-pollination treatment where, depending on the year, only 3 out of 18 seeds (16%) or none fruit (out of 18) were actually the result of self-fertilization. In 2022, pollen genotypes achieving ovule fertilization in bagged shoots of the self-pollination treatment were in fact Sikitita (7 out of 18 seeds) followed by Arbequina (5 out of 18 seeds) with Koroneiki and Tosca fathering only two and

one seeds, respectively (Table 4). A similar trend emerged in 2023 with Sikitita pollen again dominating, fathering 11 out 18 seeds where paternity could be undoubtedly assigned, followed by Arbequina with five seeds and Koroneiki with two seeds. No seeds resulted from self-fertilization in 2023 (Table 5).

**Table 4.** Number of embryos in Arbosana mother trees undoubtedly assigned to each pollen genotype (columns) in the different pollination treatments. 2022 season. LOD score threshold = 0.93. In bold, more successful fathers.

Pollination Treatment	Arbosana *	Arbequina	Sikitita	Koroneiki	Tosca	<b>Below LOD Value</b>
Self-pollination	3	5	7	2	1	7
Open-pollination	0	6	29	1	6	23
× Arbequina	0	47	4	0	3	3
$\times$ Sikitita	0	2	71	0	1	1

 $* \times$  Arbosana represents self-fertilization. Fertilization by a different genotype denotes some degree of pollen contamination in self- and cross-pollination treatments.

**Table 5.** Number of embryos in Arbosana mother trees undoubtedly assigned to each pollen genotype in the different pollination treatments during 2023 season. LOD score threshold = 0.88. In bold, more successful fathers.

<b>Pollination Treatment</b>	Arbosana *	Arbequina	Sikitita	Koroneiki	Tosca	Below LOD Value
Self-pollination	0	5	11	2	0	2
Open-pollination	0	26	27	3	5	15
×Koroneiki	0	3	0	7	2	8

 $* \times$  Arbosana represents self-fertilization. Fertilization by a different genotype denotes some degree of pollen contamination in self- and cross-pollination treatments.

Some pollen contamination, though at a much lower level, was also observed in cross-pollination treatments. For instance, in 2022, 47 out of 54 embryos of the treatment x Arbequina, where paternity was clearly assigned, were true to the intended treatment, while only seven were the result of contamination (four seeds fathered by Sikitita pollen and three by Tosca). A similarly low level of contamination occurred when using Sikitita as the pollen donor, where 71 out of 74 embryos where pollen donor was determined were correctly fathered by Sikitita with only one embryo fathered by Tosca and two by Arbequina (Table 4).

Once the compatibility between Arbosana and Arbequina and with Sikitita too was established, and given the high level of certainty provided, seed-paternity analyses were not repeated in 2023 for these two cross-pollination treatments. Therefore, in 2023, analyses focused on seeds resulting from cross-pollination treatment using Koroneiki pollen, revealing that 7 out of 12 embryos were certainly the product of fertilization by Koroneiki pollen, while three and two seeds were fathered by Arbequina and Sikitita, respectively (Table 5).

Under open pollination, Sikitita pollen was highly successful, fathering 29 out of 42 embryos with confirmed paternity in 2022. Arbequina did the same but with only six embryos, as Tosca pollen did, while Koroneiki fathered only one seed. In 2023, Arbequina and Sikitita performed equally well under open pollination, fathering of 26 and 27 seeds out of 61, respectively, while Tosca and Koroneiki were responsible for the production of five and three seeds, respectively. Notably, no seed resulted from self-fertilization in open pollination treatment across both seasons. No seeds were produced either due to fertilization by any of the many other olive genotypes growing around but out of the experimental orchard.

## 4. Discussion

Arbosana olive exhibited a clear self-incompatible response, with ISI values ranging from 0.15, compared with optimum open pollination, to 0.30 compared to cross-pollination

with Arbequina pollen. This means that Arbosana had a substantial reduction in its final fruit set (between three and six times) under self-pollination compared to cross- and/or open-pollination treatments. The differences among treatments in the initial fruit set, a parameter that measures pollinizer efficiency, were in fact substantially more pronounced than the differences found in the final fruit set across both years. This change is due to the markedly more intense fruitlet competition and subsequent abscission ("June drop") when the initial fruit set is heavier. Therefore, the differences among treatments were attenuated when the fruit population was definitively established seven weeks after bloom. The same reason applies to the significant differences in the initial fruit set between openand cross-pollination treatments in 2022 that were not statistically maintained at the final fruit set counting. Therefore, although open pollination may increase the initial fruit set with respect to cross-pollination treatments, small differences might not reach statistical significance at the final fruit set and harvest. Some differences emerged also between years with a better, higher, fruit set the second season, despite the weather being warmer and drier. Pollen adhesion and fertilization levels were also generally higher in the second year. This circumstance was likely related to the frequent rainfall during pollination in 2022; rain removes airborne pollen from the atmosphere, as the lower pollen adhesion measured in 2022 suggests. Despite the seasonal differences in fruit set under self-pollination, the self-incompatibility response was still very strong in 2023, as the ISI values show.

The self-incompatibility reaction in Arbosana is characterized by a reduced pollen tube growth in the stigma and style, rejection that leads to lower levels of ovule fertilization. While pollen adhesion on the stigma was also diminished under self-pollination, we do not interpret these differences as due to differences in stigma receptivity among treatments or due to an early rejection of self-pollen grains. The observed differences in pollen adhesion primarily stem from the procedure we used for self-pollination, specifically the constrains imposed by bagging, where the wind agitation of shoots was the sole factor facilitating pollen adhesion in self-pollinated flowers. In contrast, no bagging was performed in open pollination, while in cross-pollination treatments, a copious amount of fresh pollen was applied using a paintbrush. In any case, we do not consider the process of pollen adhesion as critical for the reduced fertilization observed under self-pollination. The value of pollen adhesion data lies in detecting that pollen adhesion was not limiting under self-pollination. Instead, the most significant parameter was the growth of the pollen tube beyond the first layers of stigmatic cells and in the style of the flowers, as previously reported by several authors [11,24–26].

Seed-paternity analyses underscore the strong rejection that Arbosana expresses toward its own pollen. Indeed, the uncontroversial paternity analyses indicate that some fruit obtained from bagged shoots were due to cross-fertilization resulting from pollen contamination. Thus, the self-incompatibility response in Arbosana was, in fact, stronger than the values suggested by the ISI values alone. This raises concerns about the procedures used to isolate the flowers and underscores the pervasive nature of olive pollen. High levels of pollen contamination have been previously reported in bagged flowers of olive and other wind-pollinated crops. For instance, Díaz et al. [8] did not find any seeds from self-fertilization in bagged branches of Picual and Arbequina nor in monovarietal orchards, leading them to conclude the strong self-incompatible condition of these cultivars. Similarly, De la Rosa et al. [27] found high levels of pollen contamination in bagged flowers intended for self-pollination and recommended installing the pollination bags in advance of bloom to prevent cross-pollen contamination in olive-breeding programs. This precaution was also advocated by Marchese et al. [7], who found no embryos fathered by Arbequina pollen in self-pollinated flowers due to unexpected contamination with foreign airborne pollen, which was likely present on the branches before bagging. Likewise, Shemer et al. [25] cautioned that viable pollen of earlier blooming cultivars might be present on the leaves of the maternal olive genotype and can be a possible source of cross-pollen contamination that can be mitigated by bagging shoots well in advance of bloom. We stress that early removal of the pollination bags is also risky and should be delayed as long as reasonably possible due to the prolonged stigma receptivity in olive [28], where even one single cross-pollen

arriving, although late, to the stigma can achieve cross-fertilization if self-incompatibility response is, as shown, robust. In contrast to the high level of pollen contamination observed under self-pollination, low levels of pollen contamination occurred in cross-pollination treatments; this is contamination that we interpret as possible when we re-opened the bags to perform hand pollinations.

Thus, our results conclusively establish Arbosana olive as self-incompatible, and therefore, it cannot be planted alone in large high-density monovarietal orchards. On the contrary, we recommend Arbosana to be inter-planted with inter-compatible cultivars to achieve optimal production in high-density orchards. Thus, identifying suitable pollinizers for Arbosana becomes imperative. In this regard, all three cultivars tested as pollinizers gave good fruit set results, confirmed by the high levels of flower fertilization, and this suggests that all can be chosen as pollen donors' trees for Arbosana. Particularly noteworthy is the effectiveness of Sikitita as the most successful pollen donor under open-pollination conditions, closely followed by Arbequina in both seasons. It is worth noting, however, that one Sikitita tree row was next to the experimental trees of Arbosana (as Tosca too, however), although in opposite wind direction of the common SW wind at the experimental site in those dates (Figure 2), while Arbequina (and Koroneiki) trees were slightly more distant but in the prevalent wind direction during bloom. Although long-distance transport of olive pollen has been documented [29–31], our study indicates that all embryos were fathered by the cultivars present in the experimental orchard at a distance between 4 and 8 m.

Both Sikitita and Arbequina emerge as highly effective pollen donors for Arbosana, suggesting their suitability as pollinizers of this cultivar. The same approach was taken for Shemer et al. [25] to recommend Picual as the best pollinizer for the cultivar Barnea. Similarly, our previous results found that the cultivars Drobnica and Lastovka were major pollen donors for the self-incompatible cultivar Oblica [24]. Seed-paternity analyses have also been instrumental in determining the best pollinizers for Kalamata, as demonstrated by Seifi et al. [18], who later extended their research to include suitable pollinizers for Barnea, Corregiola, Koroneiki and Mission [31]. However, when selecting the best pollinizers, additional criteria must be considered. Beyond the demonstrated compatibility between Arbosana and the three pollen donors under study, ideal pollinizers must bloom synchronously with the main cultivar. In this regard, in an adjacent block where the same cultivars were present, bloom phenology was determined for both seasons [32]. Figure 4 shows that some bloom overlap occurred for all cultivars, although in 2022, Sikitita showed greater overlap with Arbosana full bloom, while in 2023, Arbequina exhibited a slightly greater overlap. However, on-site assessment for extended periods is crucial to establish bloom phenology overlaps accurately. Additional criteria for selecting pollinizers for Arbosana are (i) selecting low-vigor cultivars, which makes chosen pollinizers suitable for high-density orchards, as all tested cultivars are, (ii) producing fruits having the same purpose, in this case oil production, and (iii) being regular producers, i.e., not exhibiting strong alternate bearing, in order to ensure a reliable supply of cross-pollen.



**Figure 4.** Bloom phenology in an adjacent plot to the experimental orchard in the 2022 and 2023 seasons [32].

# 5. Conclusions

In conclusion, Arbosana is a strongly self-incompatible cultivar. Its self-incompatibility response was characterized by the inhibition of pollen tube growth in the stigma and style of self-pollinated flowers, leading to significantly reduced levels of fertilization and fruit set. ISI values were consistently 0.30 or lower across both seasons, underscoring Arbosana poor performance under self-pollination compared to open and cross-pollination. Seed-paternity analyses further confirmed and quantified the minimal levels of self-fertilization observed. Given the strong self-incompatibility response of Arbosana, we discourage its cultivation in large monovarietal olive orchards. Instead, we recommend inter-planting Arbosana with trees of the cultivars Arbequina and/or Sikitita to ensure the production of high yields. For existing large monovarietal orchards of Arbosana, and while we determine the best pollination designs, we recommend re-grafting some trees with suitable pollinizers in order to mitigate the yield losses caused by its self-incompatibility. This is the first comprehensive study demonstrating the self-incompatibility response of Arbosana. Future research in this area will focus on determining the self-incompatibility response of selected low-vigor cultivars with special emphasis on the new cultivars released by different olive breeding programs.

**Supplementary Materials:** The following supporting information can be downloaded at https://www.mdpi.com/article/10.3390/horticulturae10070739/s1, Table S1: List of Arbosana seeds' embryos with LOD values below threshold (0.93), their genotype profiles, pollination treatment and replication, and likely male parent during 2022 season; Table S2: List of Arbosana seeds' embryos with LOD values below threshold (0.88), their genotype profiles, pollination treatment and replication, and likely male parent during 2023 season.

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#### References

- 1. FAO. Food and Agriculture Organization of the United Nations. Available online: https://www.fao.org/faostat/en/#data/QCL (accessed on 4 July 2024).
- Rallo, L.; Barranco, D.; Castro-García, S.; Connor, D.J.; Gómez del Campo, M.; Rallo, P. High-density olive plantations. In Horticultural Reviews; Janick, J., Ed.; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2013; Volume 41, pp. 303–383. [CrossRef]
- Connor, D.J.; Gómez-del-Campo, M.; Rousseaux, M.C.; Searles, P.S. Structure, management and productivity of hedgerow olive orchards: A review. Sci. Hortic. 2014, 169, 71–93. [CrossRef]
- García, J.M.; Hueso, A.; Gómez-del- Campo, M. Deficit irrigation during the oil synthesis period affects olive oil quality in high-density orchards (cv. Arbequina). *Agric. Water Manag.* 2020, 230, 105858. [CrossRef]
- 5. Sánchez-Estrada, A.; Cuevas, J. 'Arbequina' olive is self-incompatible. Sci. Hortic. 2018, 230, 50–55. [CrossRef]
- 6. Pinillos, V.; Cuevas, J. Open-pollination provides sufficient levels of cross-pollen in Spanish monovarietal olive orchards. *HortScience* **2009**, *44*, 499–502. [CrossRef]
- Marchese, A.; Marra, F.P.; Costa, F.; Quartararo, A.; Fretto, S.; Caruso, T. An investigation of the self- and inter-incompatibility of the olive cultivars 'Arbequina' and 'Koroneiki' in the Mediterranean climate of Sicily. *Aust. J. Crop Sci.* 2016, 10, 88–93.
- Díaz, A.; Martín, A.; Rallo, P.; Barranco, D.; De la Rosa, R. Self-incompatibility of 'Arbequina' and 'Picual' olive assessed by SSR markers. J. Am. Soc. Hortic. Sci. 2006, 131, 250–255. [CrossRef]

- 9. Baruca-Arbeiter, A.; Jakše, J.; Bandelj, D. Paternity analysis of the olive variety "Istrska Belica" and identification of pollen donors by microsatellite markers. *Sci. World J.* 2014, 208590. [CrossRef]
- 10. Breton, C.M.; Bervillé, A. New hypothesis elucidates self-incompatibility in the olive tree regarding S-alleles dominance relationships as in the sporophytic model. *C. R. Biol.* **2012**, *335*, 563–572. [CrossRef] [PubMed]
- 11. Saumitou-Laprade, P.; Vernet, P.; Vekemans, X.; Billiard, S.; Gallina, S.; Essalouh, L.; Mhaïs, A.; Moukhli, A.; El Bakkali, A.; Barcaccia, G.; et al. Elucidation of the genetic architecture of self-incompatibility in olive: Evolutionary consequences and perspectives for orchard management. *Evol. Appl.* **2017**, *10*, 867–880. [CrossRef]
- 12. De la Rosa, R.; León, L.; Guerrero, N.; Rallo, L.; Barranco, D. Preliminary results of an olive cultivar trial at high density. *Aust. J. Agric. Res.* 2007, *58*, 392–395. [CrossRef]
- 13. Larbi, A.; Ayadi, M.; Ben Dhiab, A.; Msallem, M.; Caballero, J.M. Olive cultivars suitability for high-density orchards. *Span. J. Agric. Res.* 2011, *9*, 1279–1286. [CrossRef]
- 14. Camposeo, S.; Vivaldi, G.A. Yield, harvesting efficiency and oil chemical quality of cultivars 'Arbequina' and 'Arbosana' harvested by straddle machine in two Apulian growing areas. *Acta Hortic.* **2018**, *1199*, 397–402. [CrossRef]
- 15. Polari, J.J.; Mori, M.; Wang, S.C. Virgin olive oils from super-high-density orchards in California: Impact of cultivar, harvest time, and crop season on quality and chemical composition. *Eur. J. Lipid Sci. Technol.* **2021**, *123*, 2000180. [CrossRef]
- 16. Nieto, A. Influencia de la Variedad, Densidad y Dosis de Riego en la Productividad del Olivar en seto. Master's Thesis, University of Cordoba, Córdoba, Spain, 2020.
- 17. Zapata, T.R.; Arroyo, M.T.K. Plant reproductive ecology of a secondary deciduous tropical forest in Venezuela. *Biotropica* **1978**, 10, 221–230. [CrossRef]
- 18. Seifi, E.; Guerin, J.; Kaiser, B.; Sedgley, M. Sexual compatibility of the olive cultivar 'Kalamata' assessed by paternity analysis. *Span. J. Agric. Res.* **2012**, *10*, 731–740. [CrossRef]
- 19. Sefc, K.M.; Lopes, M.S.; Mendonça, D.; Santos, M.R.; Laimer, M.; Câmara, A. Identification of microsatellite loci in olive (*Olea europaea*) and their characterization in Italian and Iberian olive trees. *Mol. Ecol.* **2000**, *9*, 1171–1173. [CrossRef] [PubMed]
- 20. De la Rosa, R.; James, C.M.; Tobutt, K.R. Isolation and characterization of polymorphic microsatellites in olive (*Olea europaea* L.) and their transferability to other genera in the Oleaceae. *Mol. Ecol. Notes* **2002**, *2*, 265–267. [CrossRef]
- 21. Carriero, F.; Fontanazza, G.; Cellini, F.; Glorio, G. Identification of simple sequence repeats (SSRs) in olive (*Olea europaea* L.). *Theor. Appl. Genet.* **2002**, *104*, 301–307. [CrossRef]
- 22. Vuletin Selak, G.; Baruca Arbeiter, A.; Cuevas, J.; Perica, S.; Pujic, P.; Raboteg Božiković, M.; Bandelj, D. Seed paternity analysis using SSR markers to assess successful pollen donors in mixed olive orchards. *Plants* **2021**, *10*, 2356. [CrossRef] [PubMed]
- 23. Kalinowski, S.T.; Taper, M.L.; Marshall, T.C. Revising how the computer program CERVUS accommodates genotyping error increases success in paternity assignment. *Mol. Ecol.* 2007, *16*, 1099–1106. [CrossRef]
- 24. Bradley, M.V.; Griggs, W.H. Morphological evidence of incompatibility in Olea europaea L. Phytomorphology 1963, 13, 141–156.
- 25. Shemer, A.; Biton, I.; Many, Y.; Vaknin, Y.; Lavee, S.; Avidan, B.; Ben-Ari, G. The olive cultivar 'Picual' is an optimal pollen donor for 'Barnea'. *Sci. Hortic.* 2014, 172, 278–284. [CrossRef]
- 26. Vuletin Selak, G.; Cuevas, J.; Goreta Ban, S.; Perica, S. Pollen tube performance in assessment of compatibility in olive (*Olea europaea* L.) cultivars. *Sci. Hortic.* **2014**, *165*, 36–43. [CrossRef]
- 27. De la Rosa, R.; James, C.M.; Tobutt, K.R. Using microsatellites for paternity testing in olive progenies. *HortScience* 2004, 39, 351–354. [CrossRef]
- Cuevas, J.; Pinillos, V.; Polito, V.S. Effective pollination period for 'Manzanillo' and 'Picual' olive trees. J. Hortic. Sci. Biotechnol. 2009, 84, 370–374. [CrossRef]
- Hernández-Ceballos, M.A.; Soares, J.; García-Mozo, H.; Sofiev, M.; Bolivar, J.P.; Galán, C. Analysis of atmospheric dispersion of olive pollen in southern Spain using SILAM and HYSPLIT models. *Aerobiologia* 2014, 30, 239–255. [CrossRef]
- García-Mozo, H.; Hernández-Ceballos, M.A.; Trigo, M.M.; Galán, C. Wind dynamics' influence on south Spain airborne olivepollen during African intrusions. *Sci. Total Environ.* 2017, 609, 1340–1348. [CrossRef]
- Mookerjee, S.; Guerin, J.; Collins, G.; Ford, C.; Sedgley, M. Paternity analysis using microsatellite markers to identify pollen donors in an olive grove. *Theor. Appl. Genet.* 2005, 111, 1174–1182. [CrossRef]
- 32. Cabello, D.; University of Córdoba, Córdoba, Spain; Cuevas, J.; University of Almería, Almería, Spain. Personal communication, 2024.

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