

1 ***In situ* ripening stages monitoring of Lamuyo pepper using a new generation NIRS**
2 **sensor**

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25 **Abstract**

26 **BACKGROUND:** Near infrared spectroscopy (NIRS) was used as a non-
27 destructive sensor to assess the quality of freshly-harvested Lamuyo peppers. 144
28 Lamuyo peppers which were in a range of colors (green, chocolate, orange and
29 red) when harvested, were analyzed. In this study the evolution of the main quality
30 parameters during the harvest period was analyzed. Additionally, NIRS predictive
31 models using a portable manual spectrophotometer to evaluate quality parameters
32 together with color index, were developed. Moreover, two procedures for taking
33 NIR spectra: 1) static, taking of point spectra readings around the equator of the
34 fruit; 2) dynamic, spectra taken by scanning the entire length of the pepper were
35 tested.

36

37 **RESULTS:** Green peppers and those harvested at the beginning of the campaign
38 presented significantly lower values ($P < 0.05$) of dry matter and soluble solid
39 contents and titratable acidity, while those with red coloration and those harvested
40 at the end of the campaign showed significantly higher values of these three quality
41 parameters ($P < 0.05$). The predictive capacity of the NIRS models showed that the
42 static mode proved to be the most suitable for measuring the quality of Lamuyo
43 peppers.

44

45 **CONCLUSIONS:** The viability of NIRS for measuring dry matter content and
46 soluble solid contents *in situ*, using a new generation NIRS sensor, was
47 demonstrated. However, the high water content, the irregular shape of the
48 vegetable and the fact that it is hollow inside, all point to the need for using larger
49 samples sets so as to increase the robustness of the models obtained.

50

51 **Keywords:** Lamuyo pepper; New generation NIRS sensor; *In situ* determination;

52 Quality parameters.

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54

55 **INTRODUCTION**

56 Lamuyo peppers are hybrids originating from France which have gradually overtaken
57 other types of peppers traditionally grown in Spain, with annual production figures of
58 around 75,000-80,000 kg/ha, due to the fact that the plants are bushy, vigorous and less
59 sensitive to cool than bell peppers. The fruits are around 12-14 cm in length and 6-8 cm
60 in equatorial diameter and are formed by 4 distinct lobes. The flesh is thick and
61 consistent, and the average weight of the fruit is 200-300 g.¹

62 Arguably, the most influential factor in the quality and postharvest life of fruit
63 and vegetable products is the degree of ripeness at the harvest time. Any fruit and
64 vegetable products harvested too early or too late in relation to their optimum state of
65 maturity are more susceptible to disease and have a shorter shelf-life than those which
66 are harvested at the best moment.²

67 Pepper fruits are considered to be in optimal condition for consumption when
68 they have attained the typical morphological characteristics of the cultivated type
69 (length, equatorial diameter and thickness of the pericarp) and have a smooth, shiny
70 skin which can be pressed without damaging the fruit. However, in peppers, the most
71 obvious aspect of the ripening process is the color change from green to red. Peppers are
72 non-climacteric fruits, and are therefore, unable to produce ethylene, the hormone
73 required for the ripening process to continue after the fruit is separated from the plant.
74 Thus, the commercial maturity of the pepper coincides with its physiological maturity,
75 and the fruits only turn red when they are on the plant.³

76 Color is, therefore, a reliable indicator of the ripeness of the pepper. In general,
77 consumers prefer dark green or bright red fruits, depending on whether they are
78 harvested before or after physiological maturity, respectively.⁴ Thus, López-Camelo and
79 Gómez⁵ suggested that the a^*/b^* ratio could be used for practical purposes as an

80 objective ripening index in peppers in order to give a realistic view of consumer
81 perceptions.

82 Therefore, for this type of vegetable, if a non-destructive quality control is set up
83 initially in the field, on the plant, this will allow to implement staggered harvesting
84 strategies in order to establish the optimum harvest time of the fruits and thus cater for
85 the demand from the industry and from consumers.

86 NIRS technology meets all the necessary requirements to be used to determine
87 the optimal harvest time for Lamuyo peppers in the field, directly on the plant itself,
88 enabling to study simultaneously various specific quality indicators of the fruit and its
89 state of maturity. At the same time, the study of the maturity curve of the pepper on the
90 plant with NIRS allows to take real-time decisions to increase production efficiency and
91 ensure product quality. All agri-food companies usually try to use these types of
92 selective harvesting strategies to adapt their production to the preferences and
93 specifications of the different markets.

94 The development in recent years of manual, portable, compact NIRS instruments
95 has enabled to make great progress in the analysis of vegetables. They can be used to
96 take agronomic decisions about the optimum harvest time as the product is growing in
97 the field, thus avoiding the need to harvest the product first and take it to the
98 laboratory.⁶⁻⁸ These advances have led to a wide range of portable instruments
99 appearing on the market, whose suitability, and that of the spectra-taking process, needs
100 to be tested before analyzing different vegetables *in situ*, such as the Lamuyo pepper.

101 The aim of this study was therefore to study the viability for measuring, in real
102 time and on the plant, the quality of outdoor-grown Lamuyo peppers of different colors
103 (i.e. at different stages of ripeness), using a manual, portable, Linear Variable Filter
104 (LVF)-based NIR instrument, which allows spectra to be taken both statically (static

105 mode), or by scanning the surface of the product (dynamic mode), in order to study the
106 evolution of different quality parameters and organize a staggered harvest, thus
107 obtaining optimum acceptance of the harvested fruits by the consumer.

108

109 **MATERIALS AND METHODS**

110 **Sampling**

111 A total of 144 Lamuyo peppers (*Capsicum annum* L.) of different colors (green = 36,
112 chocolate = 36, orange = 36 and red = 36) grown outdoors in Santaella (Córdoba, Spain)
113 were harvested manually during the months of September and October 2017.

114 The fruit was harvested first from the top of the plants and then from the lower
115 parts, since the bottom of the plant is where the peppers with a more advanced state of
116 ripeness are found (red peppers), while the unripe peppers (green peppers) are found on
117 the upper parts of the plant.

118 On arrival at the laboratory, the fruits were left at room temperature to stabilize
119 at the laboratory temperature of 20 °C.

120

121 **Spectral data acquisition**

122 The spectra were taken using the MicroNIR™ Pro 1700 (VIAVI Solutions, Inc., San
123 Jose, California, USA) instrument in reflectance mode (Fig. 1). This portable miniature
124 spectrophotometer is extremely light (64 g, not including the 150 g handle and the
125 acquisition and data processing device). The optical window measures around 227 mm².
126 This microspectrometer covers the spectral range from 910 to 1676 nm (taking data
127 every 6.2 nm), incorporating LVF. The sensor integration time was 11 ms and each
128 spectrum was the mean of 200 scans. The instrument's performance was checked every
129 10 min. A white reference measurement was obtained using a NIR reflectance standard

130 (SpectralonTM) with a 99% diffuse reflectance, while a dark reference was obtained
131 from a fixed point on the floor of the room.

132 Two modes of analysis were tested: static and dynamic. The analysis in static
133 mode was carried out thus: the sensor was placed at any given point located at the
134 equatorial diameter, at one side of the fruit, and the equipment was kept still while the
135 spectrum was being recorded; next, measurements were taken at the height of the
136 equatorial diameter, on the four sides of the fruit, rotating the fruit 90°, thus producing a
137 total of 4 measurements per fruit. The sample was placed over a black plastic sheet
138 when the spectra were taken. In the dynamic mode analysis, the four spectra taken per
139 sample were obtained thus: the sensor was moved along each of the faces of the pepper
140 analyzed, covering the area from the peduncle to the apical end of the fruit, rotating the
141 fruit 90° between measurements.

142 In both cases, the 4 spectra were averaged to provide a mean spectrum per fruit.

143

144 **Reference data**

145 To measure the a*/b* color index of the fruits, which is given by the relation between
146 the color parameters a* (red-green variation) and b* (yellow-blue variation), 4
147 measurements were taken around the fruit's equatorial diameter at 90° intervals, at the
148 same points where the NIRS spectra were taken, using a Chroma METTER CR-400
149 colorimeter (Konica Minolta Sensing INC., Osaka Japan), with illuminant C and an
150 observation angle of 2°.⁹

151 For dry matter content (DMC), 5 g of the sample was weighed on an electronic
152 scale (0-1,000 ± 0.1 g, model P1000 N, Metter-Toledo, GmbH, Greifensee,
153 Switzerland), and then dried in a hot-air oven at a temperature of 105°C until the weight

154 was constant.¹⁰ The final dry weight was calculated as a percentage of initial fresh
155 weight.

156 The soluble solid content (SSC) in °Brix was taken from the refractometer
157 reading for the pepper juice, using a temperature-compensated digital Abbe-type
158 refractometer (model B, Zeiss, Oberkochen, Würt, Germany).

159 For titratable acidity (TA), 5 g of pepper juice was used, to which 50 ml of
160 distilled water was added. Titratable acidity was measured by titration using 0.1 N
161 NaOH up to pH 8.2. The results were expressed as a percentage of citric acid.¹⁰ An
162 automatic titrator was used (Crison pH burette 24, Crison, Adella, Barcelona, Spain) to
163 take these measurements.

164 All the samples were analyzed in duplicate and the standard error of laboratory
165 (SEL) was estimated from these duplicates (Table 3). All the measurements were taken
166 immediately after the NIRS measurements.

167

168 **Data processing**

169 Data pre-processing and chemometric treatments were performed using the WinISI II
170 software package version 1.50 (Infrasoft International LLC, Port Matilda, PA, USA).¹¹

171 Before the spectral data were processed, a study was conducted to select the
172 most suitable spectral range for the instrument to carry out the quality control of
173 Lamuyo peppers. To achieve this, the 1,1,1,1 derivation treatment was applied (the first
174 digit being the number of the derivate, the second the gap over which the derivate is
175 calculated, the third the number of data points in a running average or smoothing, and
176 the fourth the second smoothing) without scatter correction, which allows to highlight
177 the areas of the spectrum where the signal/noise ratio is degraded.^{12,13}

178

179 *Spectral repeatability*

180 To calculate the spectral repeatability, the statistic Root Mean Square (RMS) was used,
181 which refers to the difference in absorbance values between several spectra taken in the
182 same sample, thus providing the mean value of the square root of the differences
183 between the spectra of a sample - in this case four - which is analyzed using the same
184 instrument, throughout the entire spectral range used.^{14,15} To establish a threshold for
185 the static and dynamic procedures, 16 Lamuyo peppers were selected, from which four
186 spectra were taken in the equatorial region both statically and by scanning the fruit's
187 surface, rotating the fruit 90° after each measurement. An admissible limit for spectrum
188 quality and repeatability was set following the procedure described by Martinez *et al.*¹⁶
189 to calculate the standard deviation limit (STD_{limit}) from the RMS statistic and obtain an
190 RMS cut- off value.

191

192 *Quantitative models: sets, calibration and validation procedures*

193 First, the structure and spectral variability of the sample population was studied to select
194 the samples that would form the sample set. To achieve this, the CENTER algorithm
195 was used, which was applied to the 144 spectra collected both statically and
196 dynamically. This algorithm performs an initial principal component analysis (PCA)
197 and determines the center of the population and the distance between each sample and
198 the center using the Mahalanobis distance (GH). Samples with a GH value over 4 were
199 considered outliers or anomalous spectra.¹⁷ As spectral pre-treatments, Standard Normal
200 Variate (SNV) and Detrending (DT) were used for scatter correction,¹⁸ together with the
201 first derivative treatment '1,5,5,1'.¹⁴

202 Once spectral outliers were removed for each mode of analysis, 111 samples
203 were selected to form part of the calibration set and the remainder (29 samples)
204 constituted the validation set (Table 2).

205 Modified partial least squares (MPLS) regression (Shenk and Westerhaus,
206 1995a) was used to obtain NIRS calibration models for the prediction of the color index,
207 (a*/b*) and quality parameters (DMC, SSC and TA) in Lamuyo peppers using the
208 MicroNIR™ Pro 1700. All regression equations were obtained using SNV + DT for
209 scatter correction¹⁸ and different derivative mathematical treatments were tested:
210 1,5,5,1; 1,10,5,1; 2,5,5,1 and 2,10,5,1.¹⁴

211 The statistics used to select the best equations with MPLS were the coefficient of
212 determination for calibration (r^2_c), the standard error of calibration (SEC), the
213 coefficient of determination for cross-validation (r^2_{cv}) and the standard error of cross-
214 validation (SECV). Furthermore, the Residual Predictive Deviation (RPD_{cv}) for cross-
215 validation was calculated as the ratio of the standard deviation (SD) of the reference
216 data to the SECV. This statistic enables SECV to be standardized, facilitating the
217 comparison of results obtained with sets of different means.¹⁹

218 Once the best predictive model was selected by statistical criteria for each
219 parameter analyzed using the two modes of analysis, tests were run for significant
220 differences between models for each parameter, with a view to identifying the most
221 suitable mode of analysis for routine use in Lamuyo peppers during the growing period
222 on the plant. The SECV values for the best equations obtained for each parameter were
223 compared using Fisher's F test.^{20,21} The values for F were calculated as:

224

$$F = \frac{(SECV_2)^2}{(SECV_1)^2}$$

225
226
227

228 where $SECV_1$ and $SECV_2$ are the standard error of prediction of two different
229 models and $SECV_1 < SECV_2$. F is compared to $F_{critical} (1-P, n_1-1, n_2-1)$ as read from the
230 table, with $P = 0.05$ and n_1 the number of times the measurement is repeated with
231 method 1; n_2 is the number of times the measurement is repeated with method 2. If F is
232 higher than $F_{critical}$, the two $SECV$ values are significantly different.

233 Finally, once the best equations for each of the two established analysis modes
234 were selected according to statistical criteria, and the best spectral sampling strategy
235 was chosen (static or dynamic mode), the models were subjected to an external
236 validation process, according to the protocol outlined by Windham *et al.*²² based on the
237 following statistics: standard error of prediction (SEP), standard error of prediction
238 corrected for bias ($SEP_{(e)}$), bias and coefficient of determination for external validation
239 (r^2_p). Generally, for calibration groups comprising 100 or more samples, and validation
240 groups containing nine or more samples, the following control limits are assumed: Limit
241 Control $SEP_{(e)} = 1.30 \times SEC$, Limit Control bias = $\pm 0.60 \times SEC$ and minimum value of
242 0.6 for r^2_p . Furthermore, the Residual Predictive Deviation (RPD_p) for prediction was
243 calculated as the ratio of the standard deviation (SD) of the validation data to the SEP.

244

245 **Statistical analysis**

246 In order to study the influence of the coloration at harvest on the DMC, SSC and TA of
247 Lamuyo pepper, a one-factor analysis of variance (ANOVA) was carried out, using
248 Statgraphics Centurion XV (StatPoint Inc., Warrenton, North Virginia, USA).

249 Next, the differences between the means were compared with the Fisher's Least
250 Significant Difference (LSD) test, and differences at $P < 0.05$ were considered to be
251 significant.

252

253 RESULTS AND DISCUSSION

254

255 Influence of the coloration at harvest on the quality of Lamuyo peppers

256 The result of the ANOVA test pointed to the existence of significant differences ($P <$
257 0.05) between the different colorations of Lamuyo peppers analyzed for the three
258 parameters tested. The results of Fisher's LSD test for DMC, SSC and TA are shown in
259 Table 1.

260 As regards DMC, green Lamuyo peppers had a significantly lower value of the
261 parameter ($P < 0.05$) than that found in chocolate-colored and red peppers, with no
262 significant differences being found in orange-colored peppers. Similarly, red peppers
263 had a significantly higher DMC ($P < 0.05$) than green and orange peppers, although no
264 significant differences were found with chocolate-colored peppers.

265 As for the SSC parameter, the green peppers presented significantly lower
266 values for this parameter ($P < 0.05$) than those found in the other colors, while red
267 Lamuyo peppers had the highest content in this parameter, which was significantly
268 higher ($P < 0.05$) than the other colors. Chocolate- and orange-colored peppers had the
269 same content in soluble solids, in between the value of the other two colors. These
270 results coincide with those of Sánchez *et al.*²³ who analyzed the dry matter and soluble
271 solid contents just after harvesting bell peppers of different colors.

272 Regarding to the TA parameter, the green peppers were the least acidic,
273 followed by chocolate, orange and lastly red peppers, which had the highest values of
274 this parameter, with significant differences ($P < 0.05$) in TA found between the different
275 colors, which represent different stages of the fruit's ripeness. These results are similar
276 to those obtained by Ghasemnezhad *et al.*²⁴ who showed an increase in the titratable
277 acidity of bell peppers throughout the ripening process, since while the fruit ripens, the

278 metabolic reactions increase, increasing the concentration of organic acids involved in
279 the Krebs cycle. For the green bell peppers, these organic acids are present in small
280 quantities, as the ripening process has not yet started.

281 It is important to note that for Lamuyo peppers, no study has been found in the
282 literature which evaluates the quality of the fruits in different stages of ripeness, as
283 reflected by their colors. Janse²⁵ however, studied the influence of the degree of
284 ripeness represented by the different colors at harvest time (green, red, yellow and
285 orange colors) in bell peppers. In that study, the author showed that dry matter and total
286 acid contents were around 25% and 60% lower, respectively, in green peppers than in
287 the other colors, and that these peppers were the least sweet, the least aromatic and had
288 the least pleasant taste. It was the red peppers that presented the highest percentages of
289 dry matter content (8.4%) and acids (3.7 mmol/100 g), the best aroma and a higher
290 content of glucose and fructose, which gave them a better flavor. Orange and yellow
291 fruits showed no significant differences between them.

292

293 **Optimum spectral region and spectral repeatability**

294 Prior to the development of the models, it was necessary to optimize the NIRS analysis
295 by means of the spectrum quality and repeatability measurement.

296 For this purpose, the existence of noise in the spectrum (spectral range 910-1676
297 nm) was evaluated. To achieve this, the derivate treatment 1,1,1,1 was applied in both
298 analysis modes in order to determine the area of the spectral range affected by noise, as
299 this it degrades the signal/noise relationship. After this process, the spectral range
300 between 1459-1676 nm was eliminated due to the high level of noise detected and all
301 the models were designed using the spectral range 910–1458 nm.

302 Spectral repeatability is crucial to the construction of models that are both
303 accurate and robust. The mean STD for the samples analyzed was 52,244 $\mu\log(1/R)$
304 (static mode) and 52,337 $\mu\log(1/R)$ (dynamic mode), representing a STD_{limit} of 65,702
305 $\mu\log(1/R)$ (static mode) and 68,131 $\mu\log(1/R)$ (dynamic mode). As can be seen, the
306 values obtained for mean STD and STD_{limit} for both modes of analysis were practically
307 identical. However, any slight differences detected in the dynamic mode could be
308 accounted for by the movement of the instrument during the NIR analysis, which could
309 cause slight deviations in the measurements.

310 When the RMS value of the each of the 4 spectra of each sample for the two
311 strategies devised did not exceed the value of the STD_{limit} , these spectra were then
312 averaged and subsequently used to perform the calibrations. In this way, a high sample
313 repeatability was achieved, which is essential for obtaining robust equations.

314

315 **Spectral features**

316 Second-derivative spectra ($D_2\log(1/R)$) for Lamuyo peppers in different stages of
317 ripeness represented by the different colors at harvest time (green, chocolate, orange and
318 red), captured by the instrument MicroNIRTM Pro 1700, together with the most relevant
319 absorption bands, are shown in Fig. 2.

320 In the NIR region between 910 and 1458 nm, absorption peaks at 978 nm, 1065
321 nm, 1120 nm, 1164 nm, 1294 nm, 1338 nm, 1369 nm and 1400 nm, mainly related with
322 C–H combination and O–H first overtone,^{26,27} appear to be especially relevant for the
323 classification of Lamuyo peppers by ripeness stage.

324

325 **Prediction of color and quality parameters using MPLS regression**

326 After using the CENTER algorithm to study the structure and spectral variability, 2 and
327 3 anomalous samples were detected in the static and dynamic modes, respectively, one
328 of which was anomalous for both strategies. Therefore 4 anomalous samples were
329 obtained for the two modes of spectral analysis, which were then removed.

330 Table 2 shows the characteristics of the calibration and validation sets used to
331 develop the predictive models for the parameters analyzed.

332 Structured selection based on spectral information, using the CENTER
333 algorithm, proved suitable, in that the calibration and validation sets displayed similar
334 values for range, mean and SD for all the study parameters; moreover, the established
335 ranges of the validation lay within those of the calibration set.

336 The calibration statistics for the best models for predicting color index and
337 quality parameters in Lamuyo peppers analyzed in static and dynamic modes are shown
338 in Table 3.

339 Regarding the color index (a^*/b^*), the predictive capacity of the model
340 developed from the spectral reading taken statically allowed to distinguish between
341 high, medium and low values for this parameter, while with the predictive capacity
342 obtained through surface scanning the product, enabled to distinguish between high and
343 low values.^{15,19}

344 No previous studies have been found in the scientific literature on measuring the
345 color index (a^*/b^*) in peppers using NIRS technology, although values for a^*/b^*
346 increase significantly during ripening due to higher carotenoid levels, thus also
347 providing a useful indicator of the fruit's ripeness.²⁸ However, Clément *et al.*²⁹ and
348 Torres *et al.*³⁰ used NIRS technology to predict the color index in tomatoes using Varian
349 Cary 500 UV-VIS-NIR (spectral range 400–1000 nm) and Perten DA-7000 (spectral
350 range 400 a 1700 nm) spectrophotometers, obtaining models whose predictive

351 capacities ($RPD_{cv} = 2.81$ and $RPD_{cv} = 2.23$, respectively) were higher than those
352 obtained here, which shows how difficult it is to take pepper color measurements during
353 the ripening process on the plant given the irregular distribution of this parameter
354 throughout ripening, as well as the convenience of using instruments which focus on the
355 visible region of the fruit.

356 To measure the quality parameters as indicated by Shenk and Westerhaus¹⁵ and
357 Williams,¹⁹ the predictive capacity of the model obtained by static analysis for DMC,
358 according to the values of the coefficient of determination for cross-validation, allows to
359 distinguish between low, medium and high values for this parameter, while in the
360 dynamic analysis of the product, the predictive capacity of the model can be considered
361 as good. Nicolaï *et al.*³¹ stated that a RPD_{cv} value of between 1.5 and 2 means that the
362 model can discriminate low from high values of the response variable. It is important to
363 note that this parameter is crucial as a measurement of ripening and is considered of
364 vital importance for the pepper industry.

365 Ignat *et al.*³² in bell peppers, using a diode array instrument (spectral range 477–
366 950 nm) reported a predictive capacity ($RPD_{cv} = 3.8$) higher than those obtained here
367 although these authors used a wider calibration set since they chose fruits picked during
368 the growing season, from the 34th day after anthesis until full ripening (88th day after
369 anthesis), and when fully grown. It should also be remembered that Lamuyo peppers
370 show very irregular shapes and it is therefore more difficult to take NIR spectra of them
371 than in bell peppers.

372 Sánchez *et al.*²³ also studied bell peppers, and analyzed them by taking spectra at
373 the fruits' equatorial diameter using a portable manual instrument based on MEMS
374 technology (MicroPhazir, spectral range 1600-2400 nm), obtaining predictive capacity

375 models ($RPD_{cv} = 1.64$) similar to those obtained here for static analysis, which were
376 slightly lower than those obtained for analysis by scanning the surface of the product.

377 For SSC, both analysis strategies enable to distinguish between high, medium
378 and low values for this parameter^{15,19} while according to Nicolai *et al.*³¹ both models
379 can discriminate between low and high values of SSC. Reid³³ pointed out the
380 importance of measuring this parameter to determine the physiological maturity of fruit
381 and vegetables.

382 Penchaiya *et al.*³⁴ used a diode array spectrophotometer (spectral range 780-
383 1690 nm) to obtain models of predictive capacity ($RPD_{cv} = 2.08$) slightly higher than
384 those of this research work. These authors used a wide range of sample attributes in the
385 calibration set, obtained by random harvesting at various stages of ripeness. Ignat *et*
386 *al.*³², using the same instrument and the same spectral range as above, obtained models
387 of predictive capacity ($RPD_{cv} = 3.9$) higher than ours; it is important to stress the greater
388 variability of the fruits used, which also affected the dry matter parameter, as mentioned
389 above.

390 Toledo-Martín *et al.*³⁵ using an instrument based on MEMS technology Phazir-
391 1018 with a 1000-1800 nm spectral range, obtained models for SSC in 14 types of
392 pepper with a predictive capacity ($RPD_{cv} = 1.7$) very similar to that obtained in this
393 work, while Sánchez *et al.*²³ for bell peppers, obtained slightly lower values for SSC
394 predictive capacity models ($RPD_{cv} = 1.65$) than those obtained in this work, using the
395 same MEMS instrument for DMC.

396 Finally, it should be noted that the predictive capacity of the models obtained for
397 TA by means of static and dynamic analysis allows to distinguish between low and high
398 values for this parameter.^{15,19} Toledo-Martín *et al.*³⁵ obtained models for TA in 14 types
399 of pepper with a predictive capacity ($RPD_{cv} = 1.4$) similar to ours. Flores *et al.*³⁶ showed

400 that the measurement of acidity-related parameters in intact fruits is notoriously
401 difficult; nonetheless, the models developed for this parameter suggested that NIRS
402 technology may be used for screening purposes.

403 Once the calibration equations for the analyzed parameters were developed for
404 each of the modes of analysis tested, the SECV statistic values obtained for each
405 parameter in the study were compared. As can be seen in Table 4, the SECV values
406 corresponding to the color index parameter obtained with the MicroNIRTM Pro 1700 in
407 static mode are significantly lower ($P < 0.05$) than when the dynamic mode is used. For
408 the rest of the parameters analyzed, no significant differences were found between the
409 SECV values for the predictive models with the two modes of analysis tested.

410 Although it could be argued that initially the dynamic mode analysis may appear
411 to result in a better fit, as it covered the whole area of the fruits analyzed and collected
412 more information about it, the fact that the existence of lobes gives the peppers an
413 irregular surface and that in the future, the analysis is likely to be carried out in the field,
414 when the product is on the plant, means that it is better to take readings on the fruit
415 statically during its development, hence the choice of this option for taking spectra in
416 the field. In addition, it would make it easier for the producers to take spectra quicker
417 and more comfortable. Thus, the static mode appears to be the most suitable for the
418 analysis of the color index and quality parameters in Lamuyo pepper *in situ*, directly on
419 the plant.

420 After comparison of both modes of analysis, and once the static mode was
421 chosen, the models obtained with this analysis mode were externally validated, using a
422 set comprising 29 samples (Fig. 3). It is important to point out that in the case of the
423 color index (a^*/b^*), 2 samples which were initially part of the validation set, were

424 eliminated before the validation procedure was carried out because they were hardly
425 represented in the calibration set with which the predictive model were finally designed.

426 The models obtained to predict the parameters DMC and SSC met all the
427 validation requirements established in the protocol established by Windham *et al.*²², as
428 well as the validation requirements in terms of the coefficient of determination for
429 prediction, r^2_p ($r^2_p > 0.6$). Both the standard error of prediction corrected for bias
430 ($SEP_{(c)}$) and the bias were within confidence limits, and so the models thus ensure an
431 accurate prediction and can be applied routinely. However, for predicting the color
432 index a^*/b^* and titratable acidity, the r^2_p and $SEP_{(c)}$ values do not comply with the
433 validation protocol,²² while in the case of bias, only the titratable acidity model
434 complies. The results therefore suggest that these NIRS models produced could
435 constitute an initial attempt to measure the maturity of Lamuyo peppers in the plant.

436 In addition, the external validation results obtained for SSC ($RPD_p = 2.1$) and
437 TA ($RPD_p = 1.3$) are higher than those obtained by Toledo-Martín *et al.*³⁵ for SSC
438 ($RPD_p = 1.8$) and TA ($RPD_p = 0.9$), respectively.

439 To evaluate the predictive ability of models in relation to the error of the
440 reference method, the SEL values were calculated (Table 3) and compared with the
441 SEPs. For the parameters a^*/b^* and DMC, the SEP values obtained were between 1-1.5
442 SEL, which means that the models developed have an excellent level of accuracy.^{19,37}
443 For SSC and TA, the SEP values obtained were 5 times higher than the SEL, so the
444 accuracy of the models obtained can be considered low.^{19,37} Nevertheless, it is important
445 to stress that all the limits and values recommended in the scientific literature and
446 mentioned above refer to other NIRS analysis conditions, i.e. using at-line instruments
447 and using pre-dried and ground samples. In this study, models were developed *in situ*
448 with a handheld portable instrument, using intact fruits with a high level of moisture. In

449 this case, the comparison with the limits indicated may be too restrictive. It is also
450 important to consider that whereas the reference values were obtained from the Lamuyo
451 pepper juice, the spectra were taken from a specific point of the fruits. For this reason, it
452 could be said that a sampling error occurred which was not included in the SEL values.

453 The regression coefficients for the best predictive models for color index
454 (a^*/b^*), DMC, SSC and TA are illustrated in Fig. 4. These regression coefficients show
455 significant importance for the region around at 980 nm, corresponding to water
456 absorption²⁶ and at around 1170–1360 nm, which correspond to the second overtone of
457 the C-H stretching bonds.²⁷ These results are similar to the ones obtained by Ignat *et*
458 *al.*³⁸ for different cultivars of bell pepper during their ripening process.

459

460 CONCLUSIONS

461 The results suggest that Lamuyo peppers harvested when green were those which
462 presented a lower content in DMC and SSC as well as lower values of TA, while those
463 picked when red had significantly higher values of the 3 parameters analyzed.

464 The findings also confirm the expectations raised that NIRS technology can
465 enable Lamuyo peppers to be harvested selectively according to their content in dry
466 matter and in soluble solids using the MicroNIRTM Pro 1700. Similarly, of the two
467 strategies for taking spectra studied, the most suitable was the NIRS analysis in the
468 statistic mode – taking of point spectra readings in the center of the surface of the
469 peppers analyzed, without the instrument moving during the measurement –, which
470 allows farmers to take spectra on peppers easily while they are growing on the plant.

471

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477

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591 carotenoid content in intact bell peppers. *Biosystem Eng* **114**:414 – 425 (2013).
592

593 **Table 1.** Quality of Lamuyo peppers according to color at harvest

Coloration	Quality parameters		
	Dry matter content (%)	Soluble solid content (°Brix)	Titrateable acidity (% citric acid)
Green	6.81 (0.70) ^(a)	5.94 (0.56) ^(a)	0.14 (0.03) ^(a)
Chocolate	7.28 (0.81) ^(b,c)	6.65 (0.93) ^(b)	0.20 (0.03) ^(b)
Orange	6.93 (0.99) ^(a,b)	6.65 (1.10) ^(b)	0.25 (0.03) ^(c)
Red	7.48 (1.05) ^(c)	7.24 (0.94) ^(c)	0.27 (0.04) ^(d)

594 Standard deviation in brackets.

595 ^{(a)-(d)}Means with different superscript letters in the same column differ significantly ($P <$
 596 0.05).

597

598

599 **Table 2.** Statistical analysis of the calibration and validation sample sets, including
 600 number of samples (N), data ranges, means and standard deviations (SD) and
 601 coefficients of variation (CV)

Parameter	Set	N	Range	Mean	SD	CV (%)
Color index (a*/b*)	Calibration	111	-0.69-2.33	0.61	1.06	173.77
	Validation	29	-0.67-1.92	0.90	0.89	98.89
Dry matter content (%)	Calibration	111	4.81-11.23	7.25	0.90	12.41
	Validation	29	5.41-9.08	6.85	0.89	12.99
Soluble solid content (°Brix)	Calibration	111	4.70- 9.30	6.74	0.97	14.39
	Validation	29	4.80-8.90	6.37	1.04	16.33
Titratable acidity (% citric acid)	Calibration	111	0.09-0.36	0.21	0.06	28.57
	Validation	29	0.10-0.34	0.22	0.06	27.27

602

603

604 **Table 3.** Calibration statistics for NIR-based models for predicting color and quality
 605 parameters in Lamuyo peppers

Parameter	Analysis mode	Math treatment	N	Range	Mean	SD	SECV	r^2_{cv}	RPD _{cv}	SEL
Color index (a*/b*)	Static	2,5,5,1	108	-0.69–2.33	0.62	1.07	0.72	0.55	1.49	0.78
	Dynamic	2,5,5,1	111	-0.69–2.33	0.61	1.06	0.88	0.32	1.20	
Dry matter content (%)	Static	1,10,5,1	107	4.81–8.92	7.24	0.78	0.48	0.63	1.63	0.37
	Dynamic	2,5,5,1	108	4.81–8.92	7.22	0.83	0.43	0.72	1.93	
Soluble solid content (°Brix)	Static	1,10,5,1	111	4.70–9.30	6.74	0.98	0.56	0.68	1.75	0.10
	Dynamic	1,5,5,1	111	4.70–9.30	6.74	0.98	0.55	0.69	1.78	
Titratable acidity (% citric acid)	Static	2,5,5,1	109	0.09–0.36	0.21	0.06	0.04	0.45	1.50	0.01
	Dynamic	2,5,5,1	108	0.09–0.33	0.21	0.06	0.05	0.37	1.20	

606 N, number of samples; SD, standard deviation; SECV, standard error of cross-validation; r^2_{cv} , coefficient
 607 of determination for cross-validation; RPD_{cv}, ratio of the SD of the original data to SECV; SEL, standard
 608 error of laboratory

609

610 **Table 4.** Comparison between SECV values obtained for the best models for predicting
 611 color index and quality parameters of Lamuyo pepper using the static and dynamic
 612 modes of analysis tested; Fisher test ($P < 0.05$)

Parameter	Static mode		Dynamic mode		F	F _{critical}
	N	SECV	N	SECV		
Color index (a*/b*)	108	0.72	111	0.88	1.49*	1.37
Dry matter content (%)	107	0.48	108	0.43	1.25	1.38
Soluble solid content (°Brix)	111	0.56	111	0.55	1.04	1.37
Titrateable acidity (% citric acid)	109	0.04	108	0.04	1.00	1.37

613 *: Significant differences ($P < 0.05$).
 614 N, number of samples; SECV, standard error of cross-validation.

615

616 **Figure 1.** Spectra acquisition procedure in Lamuyo pepper using the MicroNIR™ Pro

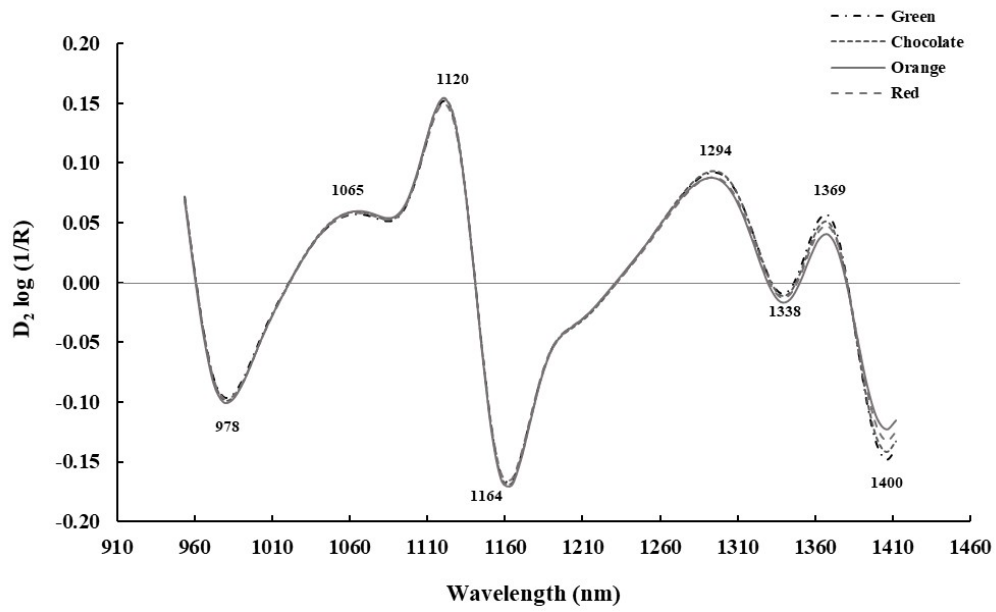
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619



620 **Figure 2.** $D_2 \log (1/R)$ spectra for Lamuyo peppers in different ripeness stages
621 represented by the different colors (green, chocolate, orange and red) at harvest time

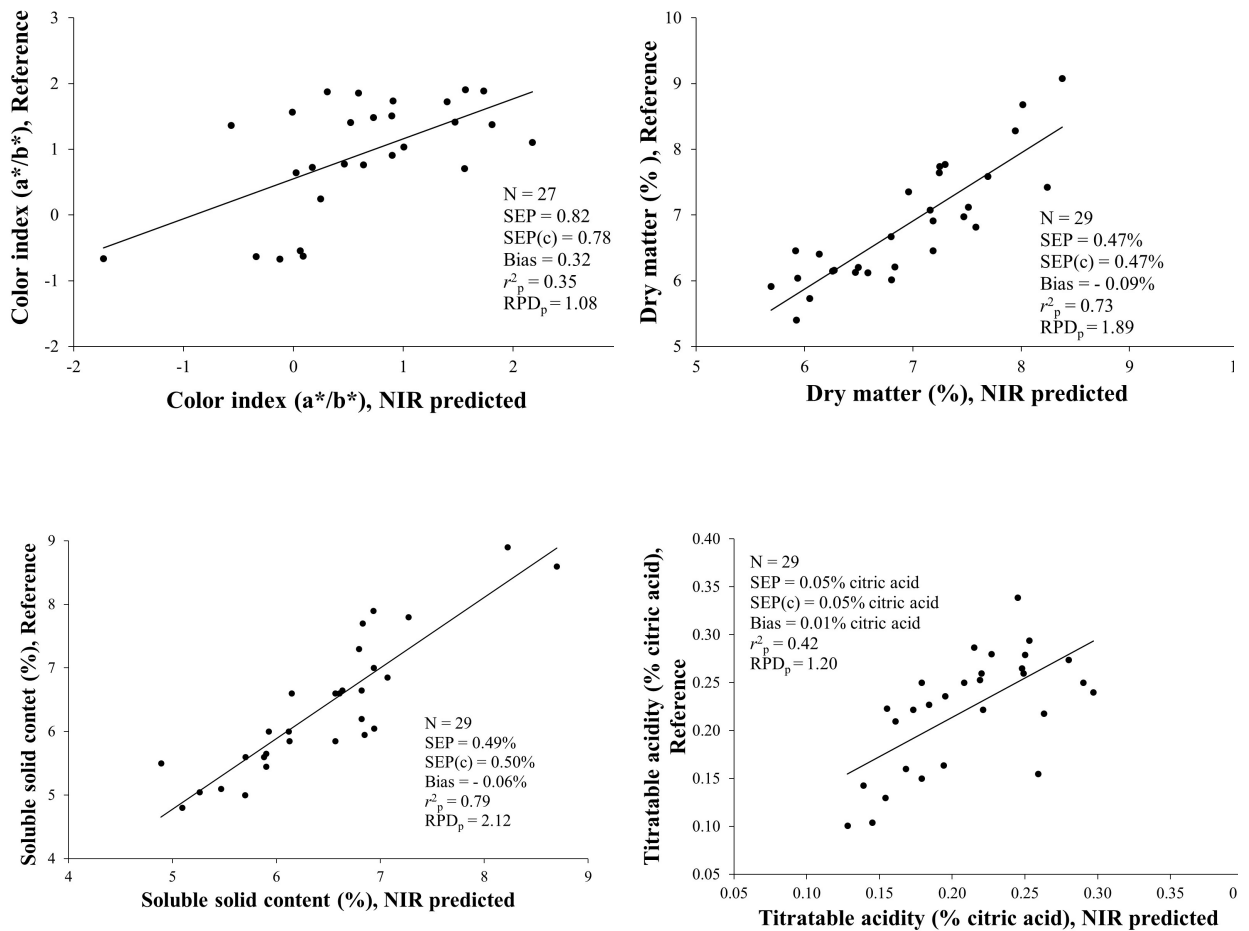


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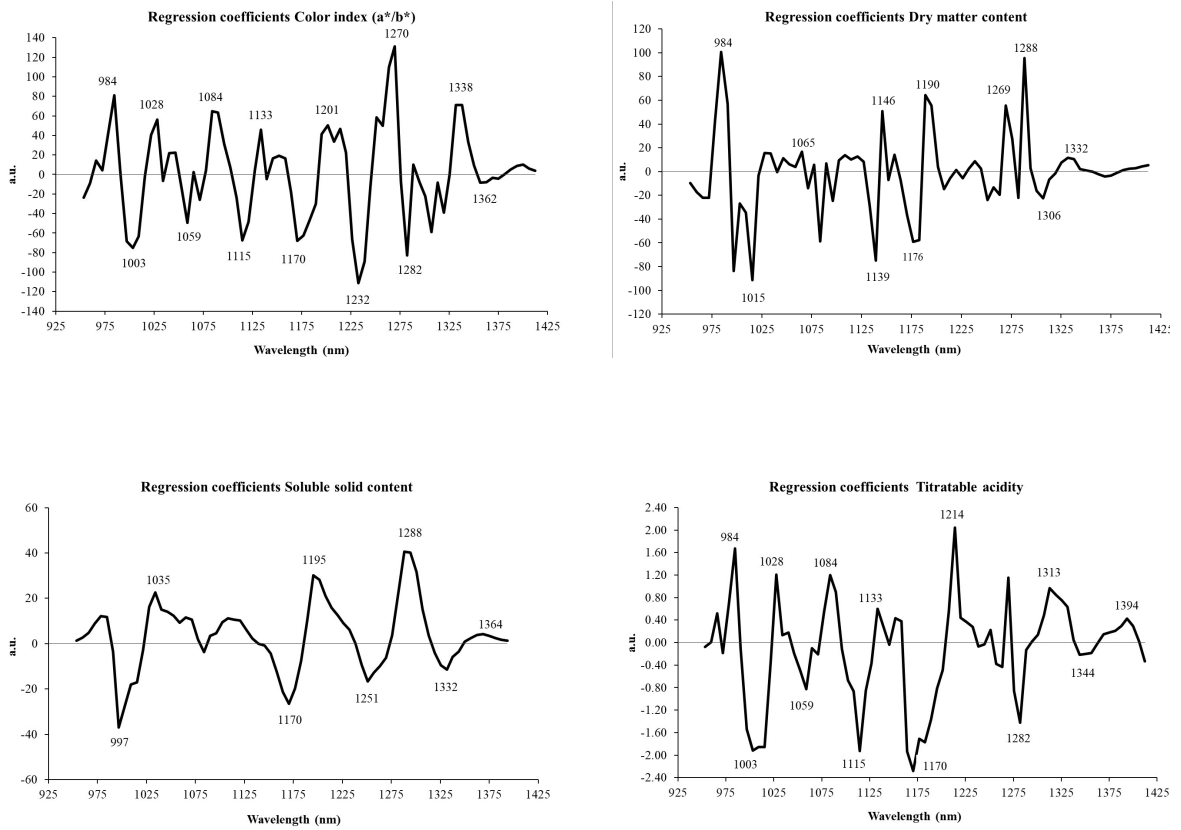
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625 **Figure 3.** Reference *versus* NIR-predicted values for color and quality parameters in
 626 Lamuyo peppers. N, number of samples for the validation set; SEP, standard error of
 627 prediction; SEP_(c), standard error of prediction corrected for bias; r^2_p , coefficient of
 628 determination for prediction; RPD_p, ratio of the SD to SEP.
 629



630

631 **Figure 4.** Regression coefficients for Lamuyo pepper color index (a^*/b^*), dry matter
 632 and soluble solid contents, and titratable acidity during on-vine ripening. * a.u.=
 633 arbitrary units
 634



635

636