

1 **Use of NIRS technology for on-vine measurement of nitrate content**
2 **and other internal quality parameters in intact summer squash for**
3 **baby food production**

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22 **Abstract**

23 This study sought to assess the feasibility of using NIR spectroscopy to predict the
24 physico-chemical composition of summer squash during on-vine ripening, with a view
25 to deciding on its possible use in baby food production depending on nitrate content at
26 harvesting. NIR calibration models were developed using a set of 157 samples scanned
27 *in situ* in the 1600–2400 nm region, using a portable handheld MEMS-NIR
28 spectrophotometer working in reflectance mode. Modified partial least squares (MPLS)
29 regression was used to interpret spectra and develop calibrations for summer squash
30 composition. Results ($r^2 = 0.83$; SECV = 112.44 mg kg⁻¹) showed that NIRS technology
31 has great potential for measuring nitrate content and also other quality parameters in
32 intact summer squashes during on-vine ripening. In addition, suitable wavelengths for
33 nitrate content determination were identified by x-loading weights and regression
34 coefficients. These findings suggest that NIRS may be a valuable tool for the rapid,
35 accurate and non-destructive measurement of nitrate content, with a view to ascertaining
36 the suitability of individual fruits for use in the production of baby foods.

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38 *Keywords:* NIR spectroscopy, summer squash, on-vine, nitrate content, baby food.

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40 **1. Introduction**

41 Over recent years, consumers have become increasingly aware of the risks
42 involved in excessive consumption of nitrates and nitrites in water and foods.
43 Vegetables are a major source of nitrates in the human diet, while nitrites are ingested
44 mainly through canned foods. In response to growing public concern, the European
45 Union passed Commission Regulation (EC) No 1881/2006 of 19 December 2006 setting
46 maximum levels for certain contaminants in foodstuffs; the maximum level for nitrates
47 in processed cereal-based foods and baby foods for infants and young children was set
48 at 200 mg NO₃/kg (OJEU, 2006).

49 Summer squash is a common ingredient in processed vegetable-based baby
50 foods. It is rich in polysaccharides, active proteins, essential amino acids, vitamins,
51 carotenoids and minerals, and provides a moderate amount of dietary fiber; interest in
52 this vegetable has increased considerably in the last few years due to its nutritional
53 properties and health benefits (Reiss et al., 2012).

54 Nitrate levels at harvesting are a key issue, particularly if the summer squash is
55 to be processed for the production of baby food. Toxicity occurs due to the conversion
56 of nitrate to nitrite, which may lead to methemoglobin due to the oxidation of Fe⁺² in
57 hemoglobin. The impaired capacity of methemoglobin to deliver oxygen to tissues may
58 lead to severe toxic effects, and may even prove fatal where methemoglobin accounts
59 for over 70% of total hemoglobin. This occurs almost exclusively in infants and very
60 young children, due to: lower stomach acidity (favoring the growth of bacteria able to
61 convert nitrate to nitrite); the presence of fetal hemoglobin (which is more easily
62 oxidized by nitrite); and lower levels of NADH-dependent methemoglobin reductase, an
63 enzyme capable of reducing methemoglobin, which is very efficient in adults
64 (Santamaria, 2006). In recent years, a number of studies have highlighted a possible link

65 between nitrate exposure and childhood type 1 insulin-dependent diabetes mellitus (van
66 Maanen et al., 2000).

67 All this has prompted greater attention to squash quality and safety concerns; as
68 a result, producers are increasingly anxious to provide consumers with assurances
69 regarding the quality and provenance of this product. Nitrate accumulation in squashes
70 depends not only on type and genetic variety, but also on a number of other factors,
71 including temperature, sunlight, available nitrogen and growing method (Blom-
72 Zandstra, 1989).

73 There is a clearly need for non-destructive sensors that can be used in the field to
74 measure squash nitrate content as well as other internal quality parameters (firmness,
75 dry matter and soluble solids content, pH and titratable acidity); on the basis of the
76 values obtained, decisions can be taken regarding optimum harvesting times and
77 possible industrial uses.

78 Near-infrared spectroscopy (NIRS), in conjunction with the application of
79 multivariate analysis strategies, is a valuable tool with great potential for the agrifood
80 sector, ensuring rapid and reliable measurement of these parameters; over recent years,
81 the field implementation of NIRS techniques has been helped by the development of
82 compact, portable instruments, which may be hand-held or tractor-mounted, and can
83 thus be readily used in the field.

84 There are no reports in the literature regarding the use of MEMS-NIRS
85 instruments for the pre-harvest monitoring of summer squashes with a view to
86 establishing the optimum time for harvesting depending on their potential destination in
87 the industry, since research to date on the use of NIRS technology for summer squash
88 quality control has focused only on the measurement of dry matter, hue angle h^* and
89 firmness using a NIR-AOTF spectrophotometer (Barnaba et al., 2012), and on the

90 determination of antioxidant compound content (Blanco-Díaz et al., 2014) and mineral
91 and carotenoid content (Martínez-Valdivieso et al., (2014a, b) using a monochromator
92 instrument to analyze lyophilized, ground product.

93 Several authors have highlighted the viability of NIRS technology for the non-
94 destructive measurement of nitrate content in various fruits and vegetables, including
95 Japanese radishes (Ito et al., 2003), leaf stalk of Qing gin cai (Ito and Idezawa, 2006),
96 spinach leaves (Xue and Yang, 2009), and pineapple (Srivichien et al., 2015).

97 This study sought to assess the feasibility of using NIR spectroscopy, with a
98 low-cost, miniaturized, handheld, near-infrared device based on MEMS technology, for
99 characterizing internal quality variations—particularly nitrate content—in intact
100 summer squashes during on-vine ripening, with a view to optimizing harvesting times
101 and enabling staggered harvesting by quality, thus allowing certain harvested squashes
102 to be used in the production of baby foods.

103 **2. Material and methods**

104 *2.1. Sampling*

105 A total of 157 summer squashes (*Cucurbita pepo* subsp. *pepo* var. *Mirza*), grown
106 on an open-air plantation in the district of La Montiel, Santaella (Córdoba, Spain),
107 were harvested between May and July 2015.

108 *2.2. Reference data*

109 Nitrate content ($\text{mg NO}_3 \text{ kg}^{-1}$) was measured following Thompson et al., (2009),
110 using an RQFlex reflectometer (Merck, Darmstadt, Germany).

111 Firmness was measured as the maximum force required to penetrate the summer
112 squashes to a puncturing depth of 10 mm using a 3-mm cylindrical tip. Summer
113 squashes were arranged with the stem-calyx axis horizontal; the first measurement was
114 made at a point on the equator, and the second after turning the fruit through 180°.

115 Texture measurements were made using a Universal Instron Texturometer (Model 3343,
116 single-column, Instron Corporation, Norwood, MA, USA), with a head speed of 0.0008
117 m/s (50 mm/min) and a 1000 N load cell.

118 Dry matter content was determined by desiccation at 105°C for 24 h (AOAC,
119 2000); final dry weight was calculated as a percentage of initial wet weight. Soluble
120 solid content (SSC, in °Brix) was measured as the refractometer reading for summer
121 squash juice, using a temperature-compensated digital Abbé-type refractometer (model
122 B, Zeiss, Oberkochen, Würt, Germany). Values for pH and titratable acidity (TA) were
123 measured using an automatic titrator (Crison Micro TT 2050, Crison, Alella, Barcelona,
124 Spain); TA was measured by titration with 0.1 mol L⁻¹ NaOH to an end point of pH 8.1.
125 Results were expressed as % citric acid.

126 *2.3. Spectral data acquisition*

127 NIR spectra of intact summer squashes were collected in reflectance mode (log
128 1/R) using a handheld micro-electromechanical system (MEMS) instrument (Phazir
129 2400, Polychromix, Inc., Wilmington, MA, USA).

130 The Phazir 2400 is an integrated near-infrared handheld analyzer that incorporates
131 all the essential components to deliver on-vine applications. The spectrophotometer
132 scans at a non-constant interval of around 8 nm (pixel resolution 8 nm, optical
133 resolution 12 nm), across the NIR wavelength range of 1600-2400 nm, with a scan time
134 per sample of 3 s. Four spectral measurements were made on each summer squash
135 whilst on the vine, at four points located 90° from each other in the equatorial region of
136 the fruit. The four spectra were averaged to provide a mean spectrum for each fruit.

137 *2.4. Data analysis: definition of calibration and validation sets*

138 Prior to carrying out NIRS calibrations, the CENTER algorithm included in the
139 WinISI II software package ver. 1.50 (Infrasoft International LLC, Port Matilda, PA,

140 USA) was applied to ensure a structured population selection based solely on spectral
141 information, for the establishment of calibration and validation sets (Shenk and
142 Westerhaus, 1991). This algorithm performs an initial principal component analysis
143 (PCA) to calculate the center of the population and the distance of samples (spectra)
144 from that center in an n-dimensional space, using the Mahalanobis distance (GH);
145 samples with a statistical value greater than 3 were considered outliers or anomalous
146 spectra.

147 The CENTER algorithm was applied in the spectral region 1600-2400 nm.
148 Mathematical treatments SNV (Standard Normal Variate) and DT (De-trending) were
149 applied for scatter correction (Barnes et al., 1989), together with the mathematical
150 derivation treatment '1,5,5,1', where the first digit is the number of the derivative, the
151 second is the gap over which the derivative is calculated, the third is the number of data
152 points in a running average or smoothing, and the fourth is the second smoothing
153 (Shenk and Westerhaus, 1995b; ISI, 2000).

154 After elimination of outlier spectra, calibration models were initially constructed
155 using all the samples available (training set, C1) for all parameters tested (Table 1).
156 After analyzing the accuracy and precision of the models obtained, new models were
157 developed for those parameters for which the best models displayed a predictive
158 capacity sufficient at least to distinguish high, medium and low values for that
159 parameter; later, these were externally validated. For this purpose, and having ordered
160 the sample set by spectral distances (from smallest to greatest distance to the center), the
161 samples forming the validation set were selected by taking one sample out of every four
162 in the initial set, although other alternatives for the selection of this set could have been
163 used. After this procedure, the calibration (C2) and validation (V) sets thus comprised
164 the samples shown in Table 1.

165 Data were subjected to chemometric treatment using the WinISI software package
166 ver. 1.50 (ISI, 2000).

167 *2.5. Data pre-processing and calibration model construction using a linear regression*
168 *strategy*

169 NIR calibration models for the prediction of quality parameters (nitrate content,
170 firmness, dry matter, SSC, pH and TA) in intact summer squashes were initially
171 constructed using the training set C1 (comprising all available samples) using modified
172 partial least squares (MPLS) regression (Shenk and Westerhaus, 1995a), with
173 subsequent cross-validation. The calibration set was partitioned into 6 groups; each
174 group was then validated using a calibration developed on the other samples; finally,
175 validation errors were combined to obtain a standard error of cross-validation (SECV).

176 A number of different pre-processing combinations were evaluated for scatter
177 correction, including SNV and DT. Additionally, a total of four derivative mathematical
178 treatments were tested: 1,5,5,1; 2,5,5,1; 1,10,5,1 and 2,10,5,1.

179 The statistics used to select the best equations were: the coefficient of
180 determination for calibration (R^2), the standard error of calibration (SEC), the
181 coefficient of determination for cross calibration (r^2), the standard error of cross
182 validation (SECV) and the coefficient of variation (CV), defined as the ratio between
183 SECV and the mean value of the reference data in the calibration set. Furthermore, the
184 Residual Predictive Deviation (RPD) was calculated as the ratio of the standard
185 deviation (SD) of the reference data to the SECV. This statistic, together with the CV,
186 enables SECV to be standardized, facilitating the comparison of results obtained with
187 sets of different means (Williams, 2001).

188 Having analyzed the predictive capacity of the models obtained using the
189 complete sample set (training set C1), new models were developed to predict the

190 selected parameters using the second calibration set (C2). The best-fitting equations
191 obtained for this new calibration set, as selected by statistical criteria, were subsequently
192 subjected to external validation following the protocols outlined by Windham et al.,
193 (1989).

194 **3. Results and Discussion**

195 *3.1. Spectral properties*

196 Mean and standard deviation spectrum for summer squash at harvesting are
197 shown in Figure 1.

198 In the wavelength region 1600-2400 nm, the major absorption peak at around
199 1920-1930 nm was mainly related to water absorption, as was the peak at around 2200
200 nm; this was to be expected, since summer squash is around 90% water (Osborne et al.,
201 1993; Williams, 2001). Osborne et al. (1993) reported peaks at around 1780 nm and
202 2310 nm related to the first sugar-related overtone. Peaks at around 1680 nm may be
203 linked to combination bands of proteins (Williams, 2001).

204 *3.2. Population characterization.*

205 Calibration (C1 and C2) and validation (V) set details, i.e. number of samples,
206 mean, range, SD, and CV for the parameters analyzed, are shown in Table 1.

207 It should be stressed that structured selection using only spectral information
208 treatment algorithms such as CENTER proved adequate and useful, since the calibration
209 and validation sets (C2 and V, respectively) displayed similar values for mean, range
210 and standard deviation for all study parameters, and ranges for the validation set lay
211 within the range recorded for the calibration set C2.

212 As Table 1 shows, the internal quality parameter displaying the greatest
213 variability was nitrate content, with a CV of 70.37% (training set C1) and 69.42%
214 (training set C2); CV for the validation set was 73.73%. Nitrogen fertilization was

215 stopped halfway through harvesting, prompting a marked drop in nitrate content
216 thereafter. The lowest variability was recorded for pH ($CV_{\text{calibration}} = 2.69\%$ and 2.85%
217 for C1 and C2, respectively; $CV_{\text{validation}} = 2.54\%$) and titratable acidity ($CV_{\text{calibration}} =$
218 11.11% and 11.25% for C1 and C2, respectively; $CV_{\text{validation}} = 8.89\%$), values for which
219 displayed little variation over the harvesting period as a whole.

220 *3.3. Prediction of nitrate content and other internal quality parameters in summer* 221 *squash on vine*

222 Results for the best models developed using training set C1 and various
223 mathematical pretreatments are shown in Table 2. Statistical criteria were used to select
224 the best model for each study parameter.

225 This study sought to determine whether harvested summer squashes could be
226 used in the production of baby foods, for which nitrate content represents a major
227 constraint. The predictive capacity of the best model for nitrate content ($r^2 = 0.83$;
228 $SECV = 112.44 \text{ mg NO}_3 \text{ kg}^{-1}$) may be considered acceptable in terms of the limits
229 recommended by Shenk and Westerhaus (1996). This result is of particular interest, in
230 that it suggests that using a low-cost, portable NIRS instrument—suitable for use in the
231 field—the industry can rapidly classify fruits as fit or unfit for baby food production on
232 the basis of nitrate content. The European Union Commission Regulation (EC) No
233 1881/2006 stipulates that summer squashes with a nitrate content of over 200 mg NO_3
234 kg^{-1} cannot be used for the production of foods for infants and young children, and both
235 producers and processors urgently require a non-destructive technique for measuring
236 nitrate content.

237 This appears to be the first published report on the use of NIRS technology to
238 measure nitrate content in summer squashes, although a number of authors have tested
239 this technology in other vegetables. Ito et al. (2003) found that NIRS in conjunction

240 with multiple linear regression enabled satisfactory determination of nitrate content in
241 Japanese radishes, although they noted that the RMSE could be improved. Ito and
242 Idezawa (2006) used NIRS and MLR algorithm to determine nitrate content in the leaf
243 stalk of Qing gin cai, reporting a good match between real and Vis-NIR-calculated
244 values for nitrate ion content for the third leaf stalk from outside and for whole nitrate
245 ion content (0.90 and 0.76, respectively). In a 2009 study of nitrate content in spinach,
246 Xue and Yang tested the best PLS model and PCR model in the spectral range 350-2500
247 nm with an independent dataset, reporting good agreement between predicted and
248 observed values, with a correlation coefficient of 0.94 for the PLS model and 0.95 for
249 the PCR model; the RMSE of prediction was 128.2 mg kg⁻¹ for the PLS model and
250 120.8 mg kg⁻¹ for the PCR model. In the only published study in fruit, Srivichien et al.
251 (2015) measured nitrate content in pineapples by Vis-NIR spectroscopy, using a
252 monochromator instrument; the results (RPD = 2.86; CV = 13.84%) were better than
253 those obtained here, perhaps because the spectral range used was 600-1200 nm.

254 The predictive capacity of the models obtained for predicting dry matter content
255 ($r^2 = 0.66$; SECV = 0.38% fw), total soluble solid content ($r^2 = 0.68$; ETVC = 0.33
256 °Brix) and pH ($r^2 = 0.57$; ETVC = 0.11)—parameters crucial for deciding the optimum
257 time for harvesting and determining the shelf-life of summer squashes—enabled high,
258 medium and low values to be distinguished (Shenk and Westerhaus, 1996).

259 Only one published study has addressed the prediction of dry matter content in
260 summer squashes using NIRS technology: Barnaba et al. (2012) used an AOTF-NIRS
261 spectrophotometer for this purpose, in conjunction with PLS regression. The predictive
262 capacity of the calibration models developed for this parameter ($r^2 = 0.81$; SECV =
263 0.46% fw) were similar to those obtained here.

264 No reports have been found in the literature regarding the determination of SSC
265 in summer squashes using NIR spectroscopy. However, Sánchez et al., (2013) measured
266 SSC in intact mandarins using a Phazir 2400 instrument in the 1600-2400 nm spectral
267 region, reporting results (RPD = 1.49; CV = 6.06%) very similar to those obtained in
268 the present study.

269 Portable MEMS-based NIRS instruments have not hitherto been used to
270 determine pH in summer squashes, but Sánchez et al. (2013) report poorer results for
271 pH prediction in intact mandarins (RPD = 1.11; CV = 2.59%), highlighting the fact that
272 pH is difficult to predict if training sets are insufficiently varied.

273 The predictive capacity of the models developed to predict firmness ($r^2 = 0.45$;
274 SECV = 1.46 N) and titratable acidity ($r^2 = 0.44$; SECV = 0.01 % citric acid) enabled
275 high and low values to be distinguished for these parameters, thus meeting the criterion
276 recommended by Shenk and Westerhaus (1996).

277 The results obtained for firmness underline the difficulty in correlating a
278 destructive measurement made at a puncturing depth of 10 mm with a non-destructive
279 measurement; as Peirs et al. (2002) have noted, NIRS light only penetrates to a useful
280 depth of between 1 and 5 mm, depending on the wavelength, the instrument used and
281 the maturity of the fruit tested.

282 The predictive capacity of the models developed to predict titratable acidity
283 reflects the fact that all summer squashes were harvested at commercial maturity, and
284 thus displayed very uniform acidity values. González-Caballero et al. (2010) report that
285 this parameter cannot be predicted using NIRS technology and MPLS regression if the
286 training sets have low standard-deviation values, as was the case here.

287 Portable MEMS-based NIRS instruments have not been used to date for the
288 prediction of firmness in summer squashes, but Pérez-Marín et al. (2010) reported a

289 predictive capacity for firmness in intact plums (RPD = 1.18; CV = 53.10%) slightly
290 lower than that obtained here, using the same instrument. This highlights the difficulty
291 of correlating measurements when using an instrument working in the 1600-2400 nm
292 spectral region.

293 Although there are no reports in the literature regarding the *in situ* measurement
294 of titratable acidity in outdoor-grown summer squashes, Sánchez et al. (2013), studying
295 quality measurements in on-tree mandarins using the Phazir 2400 instrument, obtained
296 models whose predictive capacity (RPD = 1.68; CV = 11.93%) was better than that
297 obtained here, perhaps because the training sets contained greater variability, since
298 measurements were made throughout ripening.

299 *3.4. New calibration process and external validation*

300 Once the predictive capacity of the models using the training set C1 and cross-
301 validation had been analyzed, only those models (nitrate content, dry matter, SSC and
302 pH) for which $r^2 > 0.5$ were subjected to external validation.

303 The aim was, in the first instance, to validate the best calibration models using a
304 sample set not included in the calibration, but similar to the calibration set. Validations
305 of the best calibration models obtained with training set C2 were performed using a set
306 comprising 34 samples (Table 3).

307 Models constructed for predicting nitrate content in intact summer squash using
308 the MEMS instrument met the validation requirements in terms of r^2 ($r^2 > 0.6$), while the
309 SEP(c), bias and slope were within confidence limits: the equation thus ensures accurate
310 prediction, and can be applied routinely (Windham et al., 1989). As Table 3 shows, two
311 samples in the initial validation set (nitrate content of 1,030 and 1,065 mg kg⁻¹) were
312 eliminated since the samples were unrepresentative of the calibration set (Fig. 2), thus
313 hindering their correct prediction.

314 Using the monitoring procedure, the prediction statistic values obtained for dry
315 matter fell short of the limit recommended for routine application ($r^2 > 0.60$). However,
316 it should be stressed that the SEP(c) and slope values were close to confidence limits
317 and bias was below confidence limits, suggesting that the NIRS equation for this
318 internal parameter can be regarded as a useful preliminary trial for obtaining accurate
319 on-vine quality predictions for intact summer squashes. Likewise, the values for bias in
320 the models constructed for predicting SSC and pH in intact squashes using a handheld
321 MEMS spectrophotometer lay within confidence limits, although r^2 , SEP(c) and slope
322 results did not always attain recommended minimum values, indicating that the NIRS
323 equations constructed may be considered as a first step in the fine-tuning of NIRS
324 technology for the on-vine monitoring of internal quality parameters in summer
325 squashes.

326 These results highlight the importance not only of ensuring a sufficient number
327 of samples in the calibration set, but also of guaranteeing the adequate distribution and
328 structure of the sample set.

329 *3.5. Analysis of sensitive wavelengths for the prediction of nitrate content*

330 The x-loading and regression coefficient plots for the best model obtained for
331 predicting nitrate content in intact summer squash are shown in Fig. 3. These plots show
332 the areas across the spectral range in which variance has influenced the computing of
333 the model to a greater or lesser degree, and the direction (positive or negative).

334 For the prediction of nitrate content using the Phazir 2400, representation of the
335 six latent variables (LV) used in constructing the calibration equation and the regression
336 coefficients shows that the areas of the spectrum exerting greatest weight on model
337 fitting were 1712, 1776, 1850, 1920, 1984, 2008, 2100, 2152 and 2264 nm. Their

338 influence was either positive or negative, depending on the latent variable in question
339 (Fig. 3).

340 **4. Conclusions**

341 These results suggest that NIRS is a very promising and useful sensor for the
342 non-destructive quantification of changes in nitrate content and other internal quality
343 parameters in summer squashes in the course of on-vine ripening, enabling decisions to
344 be taken regarding the optimum harvesting time. Harvested fruit can thus be swiftly
345 streamed, allowing batches with different nitrate contents to be processed separately.

346 Findings also show that the use of portable MEMS-based NIRS instruments
347 enables nitrate content to be measured, rapidly and *in situ*, during on-vine ripening, thus
348 providing the industry with a means of establishing the final destination of the product,
349 since if nitrate content exceeds the maximum levels stipulated under current legislation,
350 the fruits cannot be used in the production of baby foods.

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445

446 **Table 1.**

447 Number of samples (N), range, mean, standard deviation (SD), and coefficient of variation (CV) for different calibration sets (C1 and C2) and for
448 the validation set (V)

	Parameters																	
	Nitrates (mg kg ⁻¹)			Firmness (N)			Dry matter (% fw)			SSC (°Brix)			pH			Titratable acidity (% citric acid)		
	C1	C2	V	C1	C2	V	C1	C2	V	C1	C2	V	C1	C2	V	C1	C2	V
N	150	116	34	150	116	34	150	116	34	150	116	34	150	116	34	150	116	34
Range	30.00- 1074.00	30.00- 1074.00	55.00- 1068.00	0.25- 9.81	0.25- 9.81	1.35- 9.20	3.48- 6.74	3.48- 6.74	3.71- 5.83	3.60- 6.70	3.60- 6.70	3.70- 6.35	5.86- 6.76	5.86- 6.76	5.99- 6.60	0.07- 0.11	0.07- 0.11	0.07- 0.10
Mean	410.25	403.49	433.34	5.76	5.67	6.05	4.87	4.92	4.67	4.73	4.75	4.65	6.31	6.31	6.29	0.09	0.08	0.09
SD	288.68	280.13	69.43	2.09	2.14	1.94	0.67	0.68	0.62	0.61	0.60	0.66	0.17	0.18	0.16	0.01	0.009	0.008
CV	70.37	69.42	73.73	36.28	37.74	32.07	13.76	13.82	13.28	12.90	12.63	14.19	2.69	2.85	2.54	11.11	11.25	8.89

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450

451 **Table 2.**

452 Calibration statistics for NIR-based models for predicting internal quality parameters in

453 intact summer squash. Training set C1.

Parameter	Math treatment	N	Range	Mean	SD	SECV	r ²	CV (%)	RPD
Nitrates (mg kg ⁻¹)	2,5,5,1	139	30.00-1074.00	386.64	271.82	112.44	0.83	29.08	2.42
Firmness (N)	2,10,5,1	145	1.30-9.81	5.88	1.98	1.46	0.45	24.83	1.36
Dry matter (% fw)	2,10,5,1	145	3.48-6.74	4.84	0.65	0.38	0.66	7.85	1.71
SSC (°Brix)	1,5,5,1	144	3.70-6.00	4.71	0.58	0.33	0.68	7.01	1.76
pH	1,5,5,1	144	5.86-6.62	6.30	0.17	0.11	0.57	1.75	1.55
Titrateable acidity (% citric acid)	2,5,5,1	148	0.07-0.11	0.09	0.01	0.01	0.44	7.54	1.35

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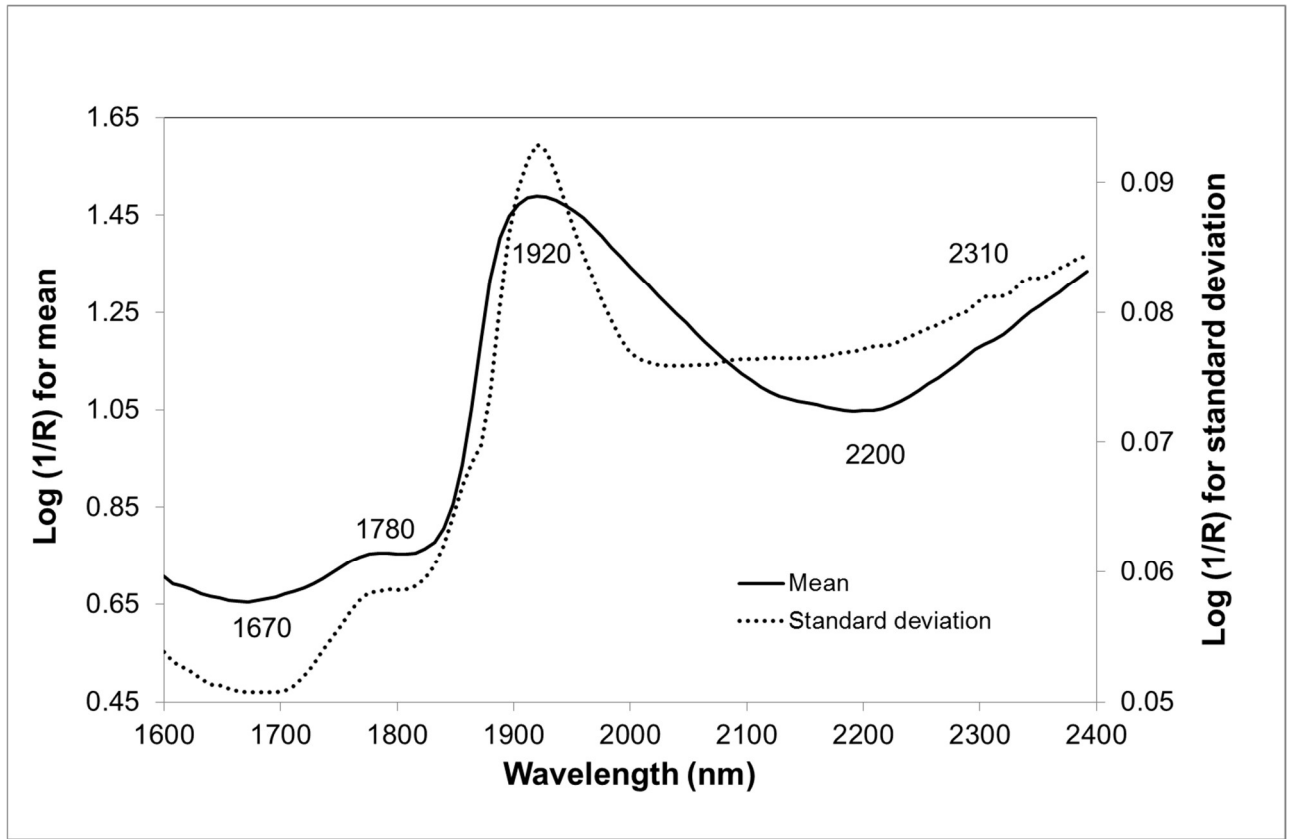
456

457 **Table 3**

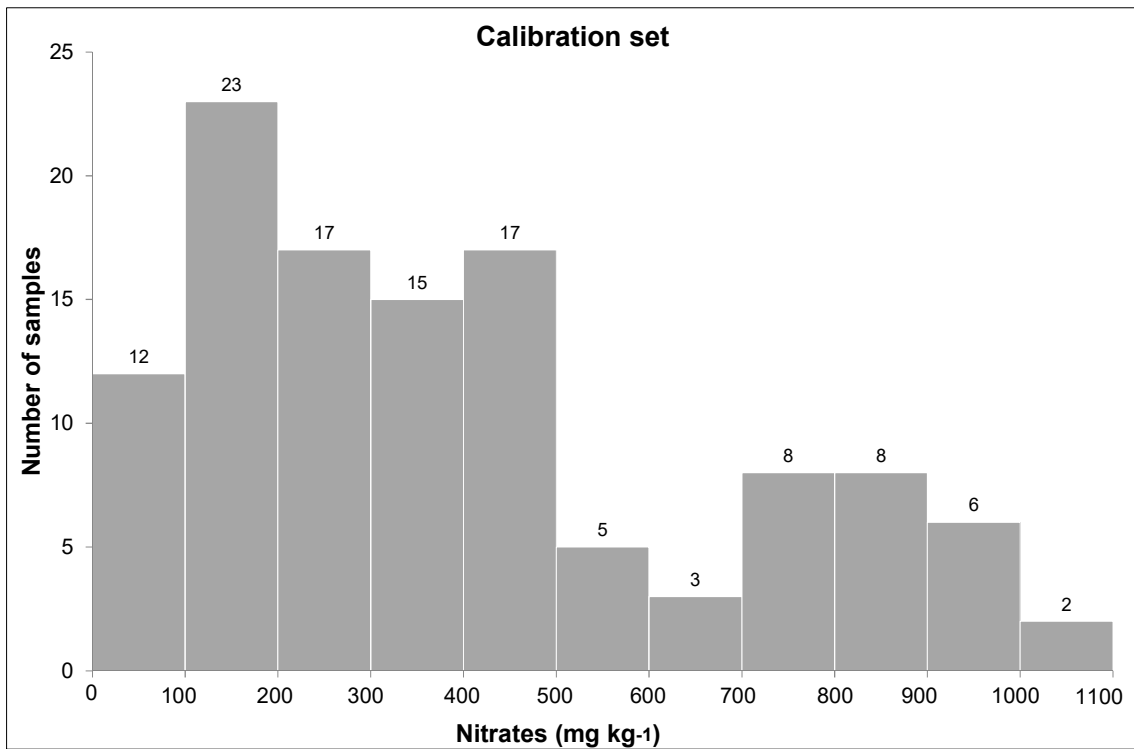
458 MPLS regression statistics for NIR-based models for predicting internal quality parameters in intact summer squash.

Parameter	Math treatment	Calibration								Validation				
		N	Range	Mean	SD	SECV	r ²	CV (%)	RPD	N	r ²	SEP	SEP (c)	Bias
Nitrates (mg kg ⁻¹)	2,5,5,1	115	30.00-1074.00	398.82	276.8	145.04	0.73	36.37	1.91	32	0.67	163.75	165.55	16.25
Dry matter (% fw)	1,5,5,1	111	3.48-6.74	4.90	0.66	0.38	0.67	7.69	1.75	34	0.50	0.48	0.47	-0.09
SSC (°Brix)	1,10,5,1	107	3.70-6.00	4.73	0.50	0.32	0.59	6.82	1.56	34	0.35	0.53	0.54	-0.02
pH	2,5,5,1	113	5.86-6.62	6.31	0.17	0.11	0.56	1.81	1.50	34	0.33	0.14	0.14	-0.01

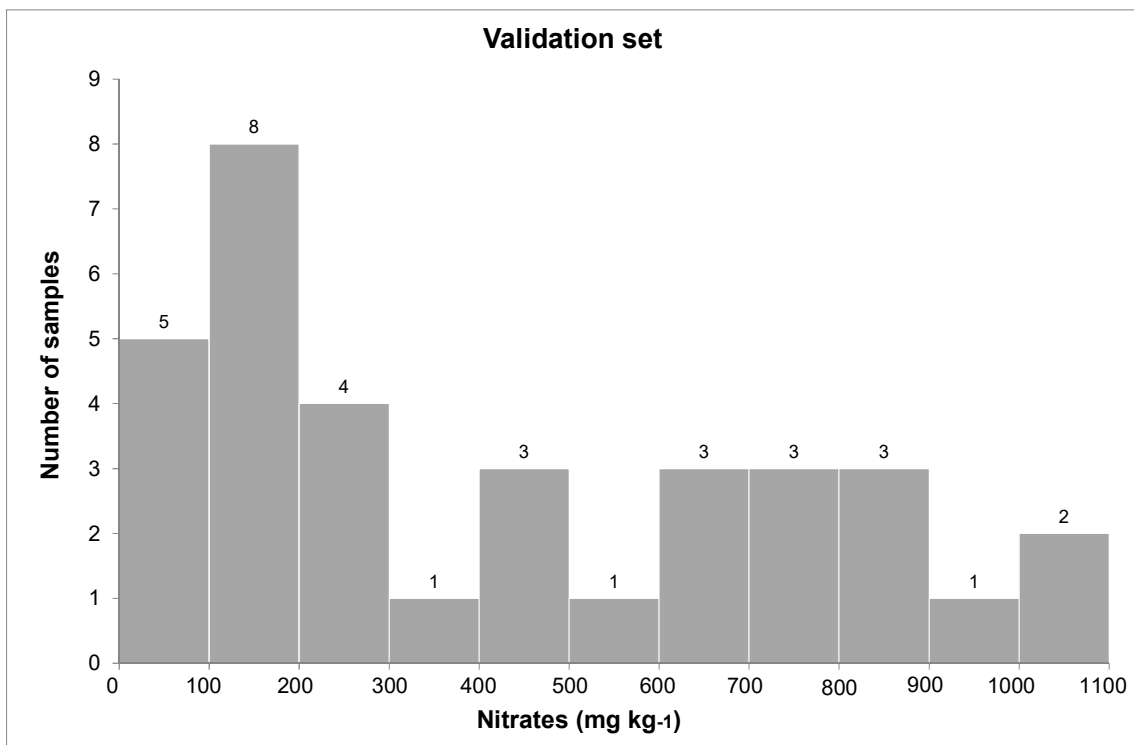
Fig. 1. Mean and standard deviation spectrum for summer squash



462 **Fig. 2.** Distribution of nitrate content for intact summer squashes during on-vine
463 ripening.



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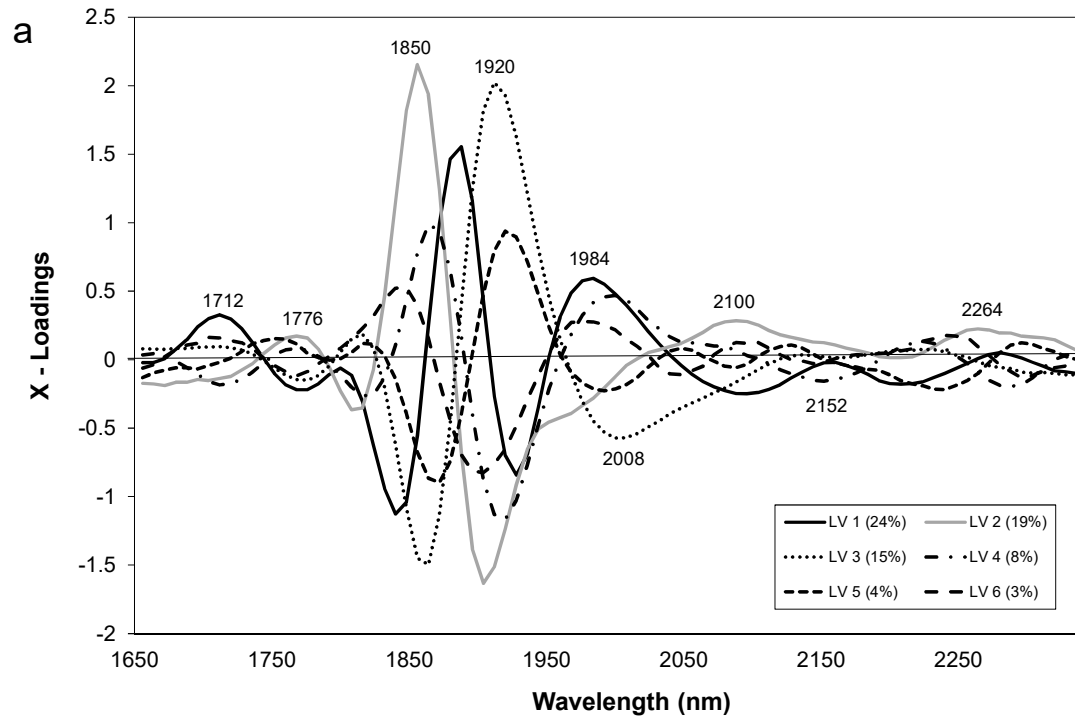
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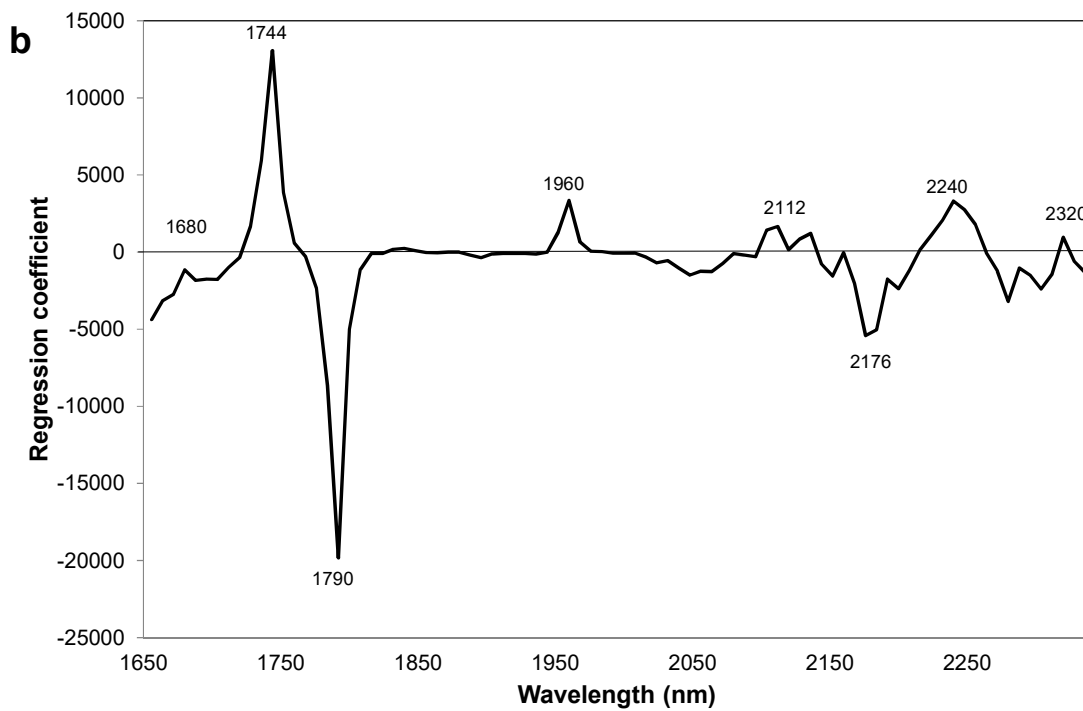
468 **Fig. 3.** x-Loading weights (a) and regression coefficients (b) for summer squash nitrate
469 content during on-vine ripening.

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